

Deciphering the molecular and cellular mechanisms underlying PFAS-associated MASLD

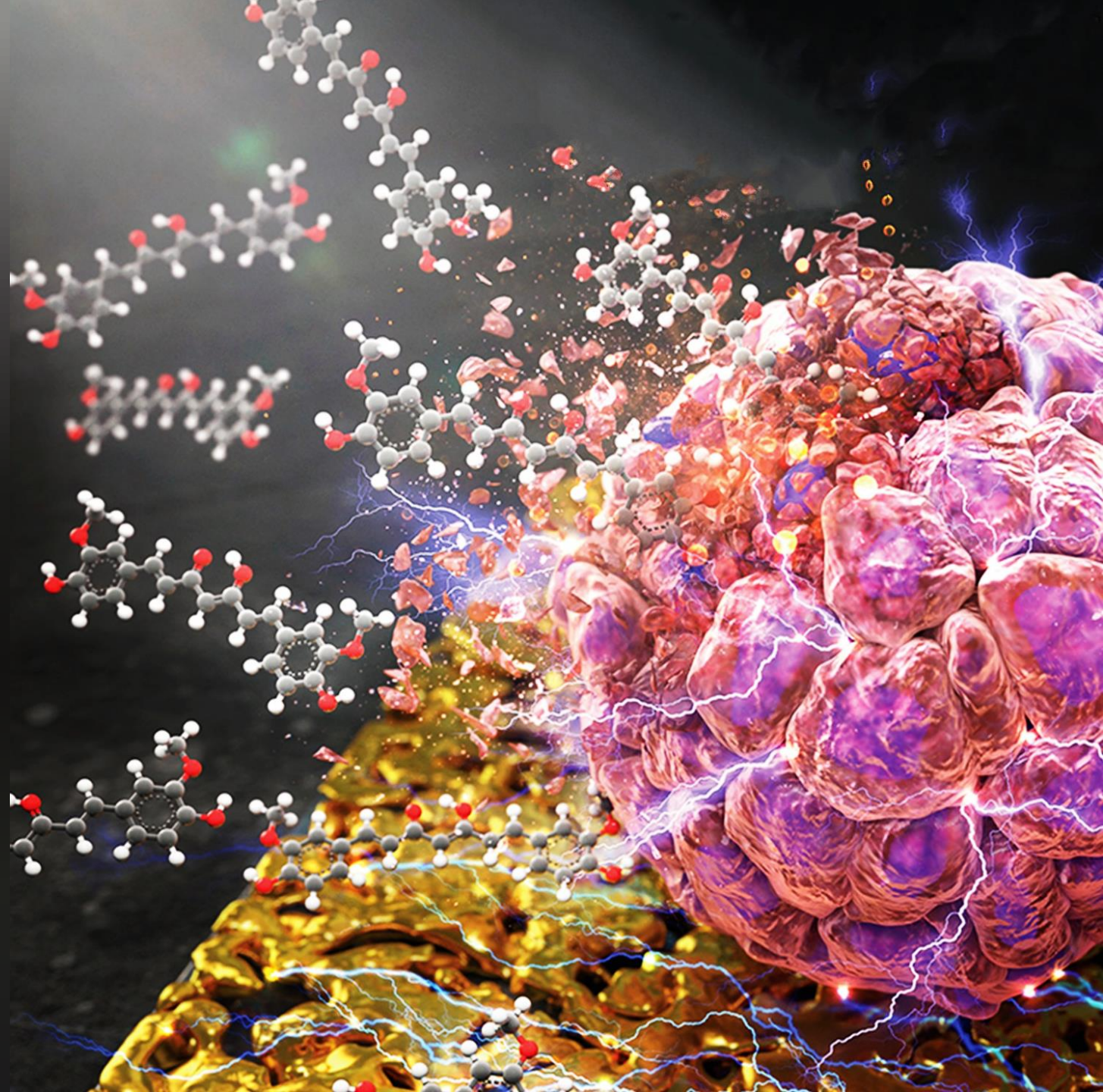
Ana Maretti-Garcia, MS, PhD

Assistant Professor, Department of Medicine,
University of Southern California

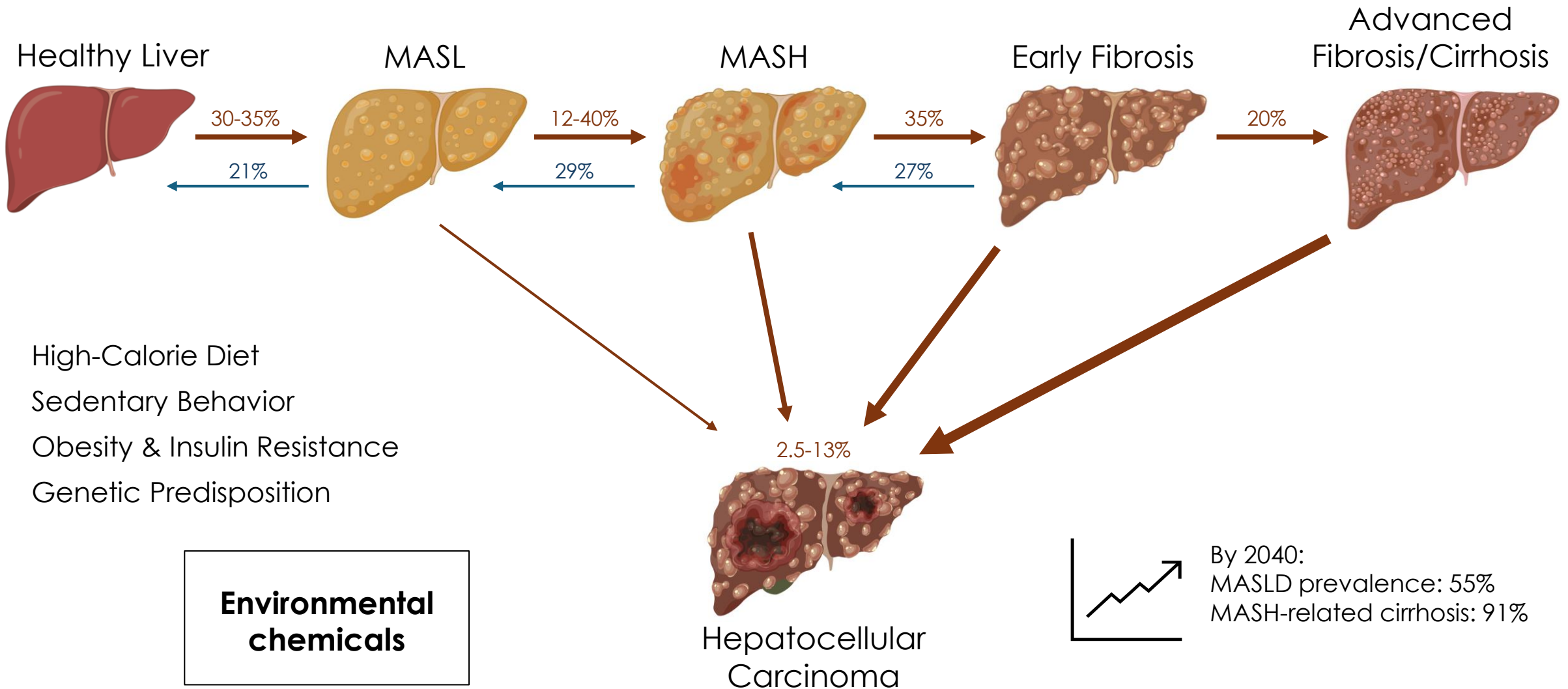
Co-Investigator, ShARP Center

**NIEHS SRP Risk e-Learning Webinar Series:
*From Cells to Solutions – Session III***

January 21, 2026



Natural History of Metabolic Dysfunction-Associated Steatotic Liver Disease (MASLD)



What are PFAS?

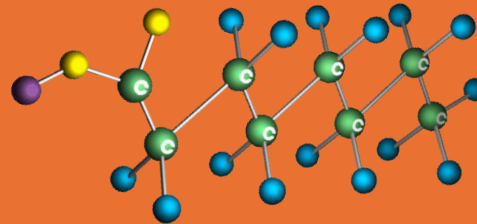
> 10,000
chemicals

Widely used in
industrial applications

Resistant to
degradation

Detected in the
blood of almost
everyone in the U.S

PFAS



Exposure Sources



Contaminated water



Nonstick cookware



Food packaging

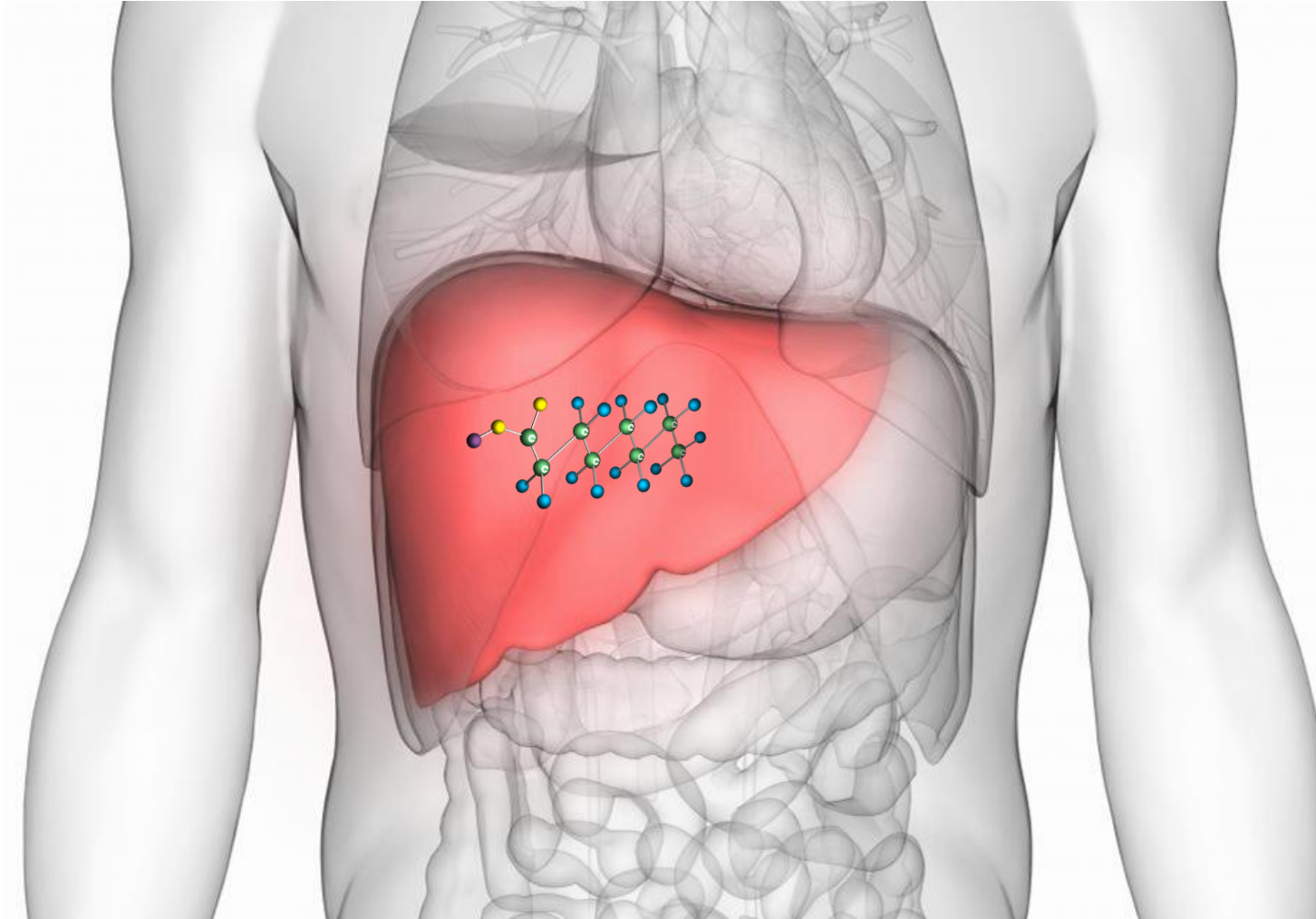


Personal care products



Textiles (Outdoor Gear &
Waterproof Clothing)

Hepatic Effects of PFAS Exposure



What do we know?

Hepatic Injury

Lin et al., 2010 (PFOA)

Gleason et al., 2015 (PFOS)

Costello et al., 2022 (PFOA, PFOS, PFNA)

Metabolic Dysfunction

Fleisch et al., 2017 (PFOA, PFOS, PFNA, PFHxS, PFDeA)

Rosen et al., 2022 (PFNA, PFOS)

Dyslipidemia

Schlezingner et al., 2021 (PFOA)

Averina et al., 2021 (PFNA, PFDA)

Steatosis and Inflammation

Jin et al., 2020 (PFOS, PFHxS)

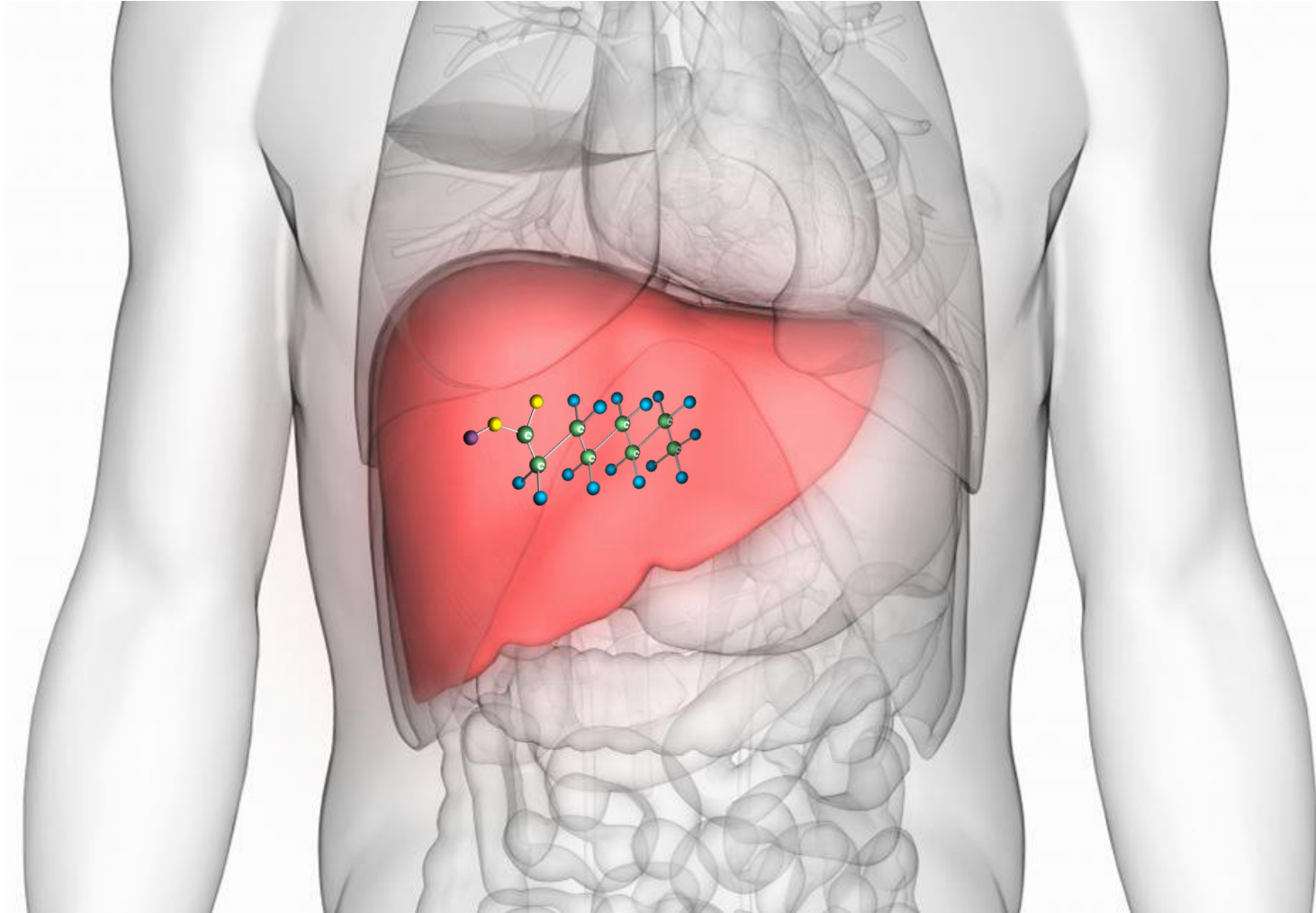
Zhang et al., 2023 (PFHxS)

Wu et al., 2023 (PFOA, PFNA)

Cancer

Goodrich et al., 2022 (PFOS)

Hepatic Effects of PFAS Exposure



Current Knowledge Gaps and Limitations

Mechanisms are unknown

- Pathways and metabolism cascades
- Cell-specific impact
- Changes in cell-cell communication

Available experimental models

- Animal models do not recapitulate human disease
- Human studies require years of follow-up
- *2D in vitro* models only use single cell types in self-assembled monolayers

Data integration

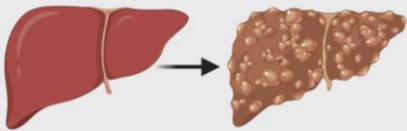
- Current studies do not integrate *in vitro* or *in vivo* datasets with human data



ShARP

SUPERFUND PFAS ASSESSMENT,
REMEDATION & PREVENTION

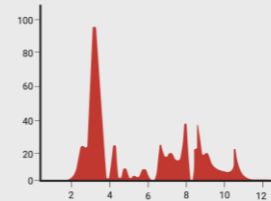
Understanding the
causal effects of PFAS
in human liver disease



Discovering
mechanisms of PFAS
hepatotoxicity



Developing methods
for comprehensive
PFAS environmental
assessment



Identifying sustainable
PFAS interventions



Biomedical Research Projects

Project 1: Mechanisms of PFAS Hepatotoxicity

Project 2: PFAS-induced MASLD in Youth

Environmental Engineering Research Projects

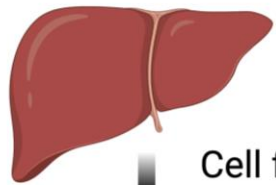
Project 3: Fate and Transport of PFAS

Project 4: PFAS Remediation and Treatment

In Vitro Study: Multi-donor 3D Human Liver Spheroids

insphero

Cell Sources

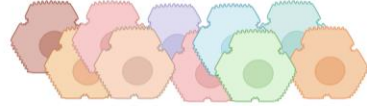


Cell from human postmortem liver

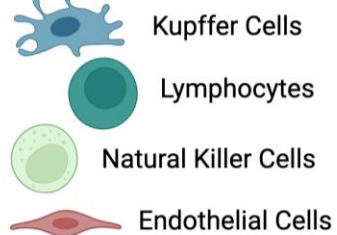


Production of 3D InSight™ Human Liver Microtissues

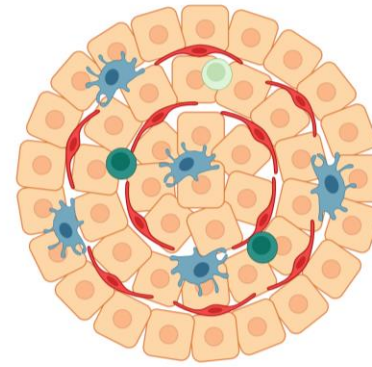
Hepatocytes
(5 male and 5 female donors)



Non-parenchymal cells
(1 donor)

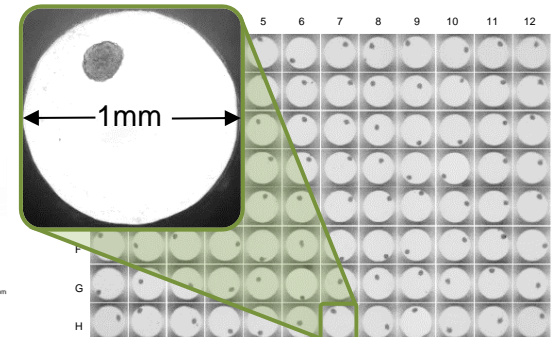
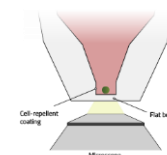


Cell aggregation





Quality Control

- Macroscopic observation (spheroid formation)
- Microtissue size
- Immunohistochemistry markers
- Cytokines
- Metabolites
- Liver enzymes






In Vitro Study: Previous PFAS toxicological studies

High-Throughput Transcriptomic Analysis of Human Primary Hepatocyte Spheroids Exposed to Per- and Polyfluoroalkyl Substances as a Platform for Relative Potency Characterization

Andrea Rowan-Carroll,* Anthony Reardon ,* Karen Leingartner,* Remi Gagné,* Andrew Williams,* Matthew J. Meier,* Byron Kuo,* Julie Bourdon-Lacombe,[†] Ivy Moffat,[†] Richard Carrier,[†] Andy Nong,* Luigi Lorusso,[‡] Stephen S. Ferguson ,[§] Ella Atlas,^{*,1} and Carole Yauk^{*,1,1}

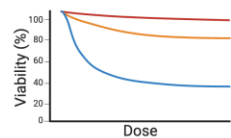
TOXICOLOGICAL SCIENCES, 181(2), 2021, 199–214

Potency Ranking of Per- and Polyfluoroalkyl Substances Using High-Throughput Transcriptomic Analysis of Human Liver Spheroids

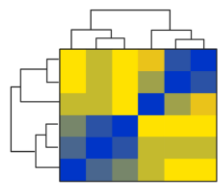
Anthony J.F. Reardon ,* Andrea Rowan-Carroll,* Stephen S. Ferguson ,[†] Karen Leingartner,* Remi Gagne,* Byron Kuo,* Andrew Williams,* Luigi Lorusso,[‡] Julie A. Bourdon-Lacombe,[§] Richard Carrier,[§] Ivy Moffat,[§] Carole L. Yauk,^{*,1,1} and Ella Atlas ,^{*,1,1}

TOXICOLOGICAL SCIENCES, 184(1), 2021, 154–169

Cytotoxicity Assessment



LDH assay



TempO-Seq

Table 1. Number of Differentially Expressed Genes in the PFAS-Treated Spheroids Relative to Controls for Each Time Point and Concentration

PFAS	Day	0.02 μ M	0.1 μ M	0.2 μ M	1 μ M	2 μ M	10 μ M	20 μ M	50 μ M	100 μ M
PFOA	1	0	8	36	8	19	79	69	227	465
	4	22	30	35	41	14	42	100	184	782
	10	14	7	2	4	10	82	101	593	Cytotoxic
	14	4	0	8	9	6	76	96	Cytotoxic	Cytotoxic
PFBS	1	0	1	0	5	49	5	0	44	73
	4	0	5	20	20	0	0	8	0	59
	10	2	1	0	1	2	0	7	54	76
	14	0	0	6	0	1	2	0	2	52
PFOS	1	1	85	3	6	51	167	277	Cytotoxic	Cytotoxic
	4	6	12	30	48	62	251	313	Cytotoxic	Cytotoxic
	10	2	7	4	6	60	163	466	Cytotoxic	Cytotoxic
	14	1	0	3	1	6	249	398	Cytotoxic	Cytotoxic
PFDS	1	0	1	0	6	22	59	81	177	186
	4	0	17	15	7	20	340	247	171	273
	10	0	30	0	43	40	134	175	232	231
	14	1	3	1	18	100	182	147	211	340

In Vitro Study: PFAS impact in Liver Spheroids



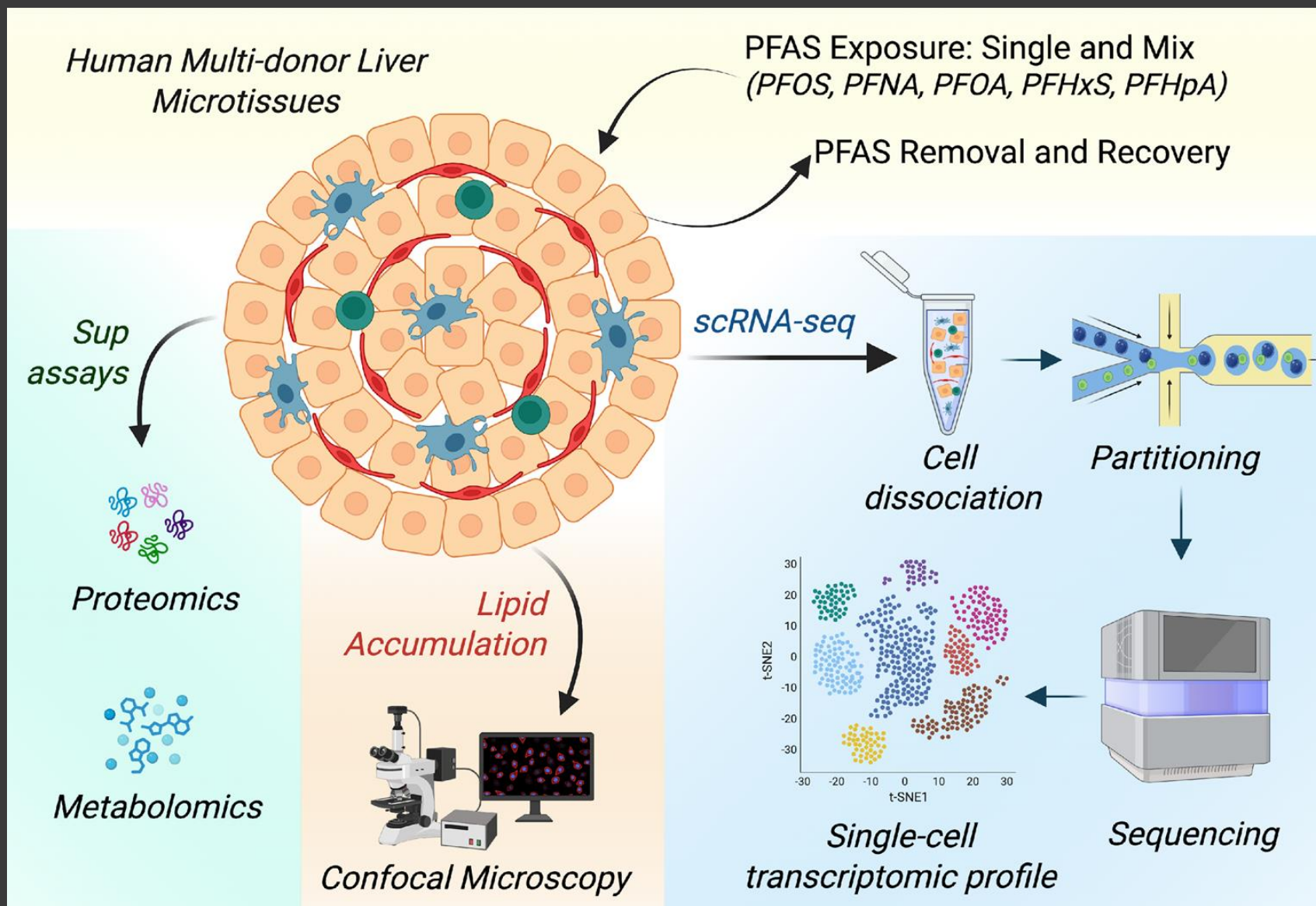
Lucy Golden, PhD
PI -Project 1



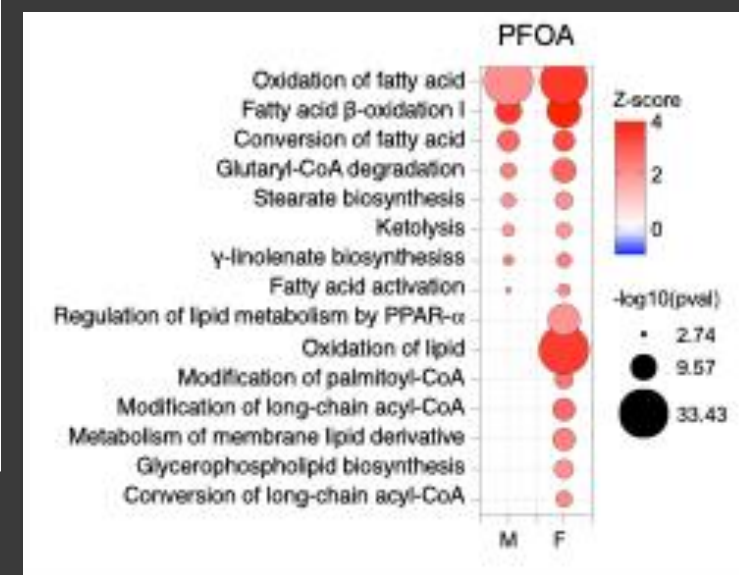
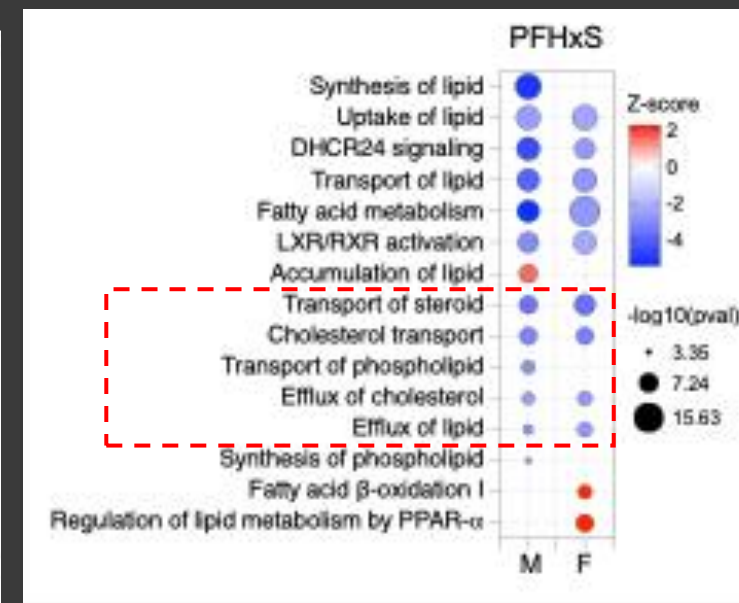
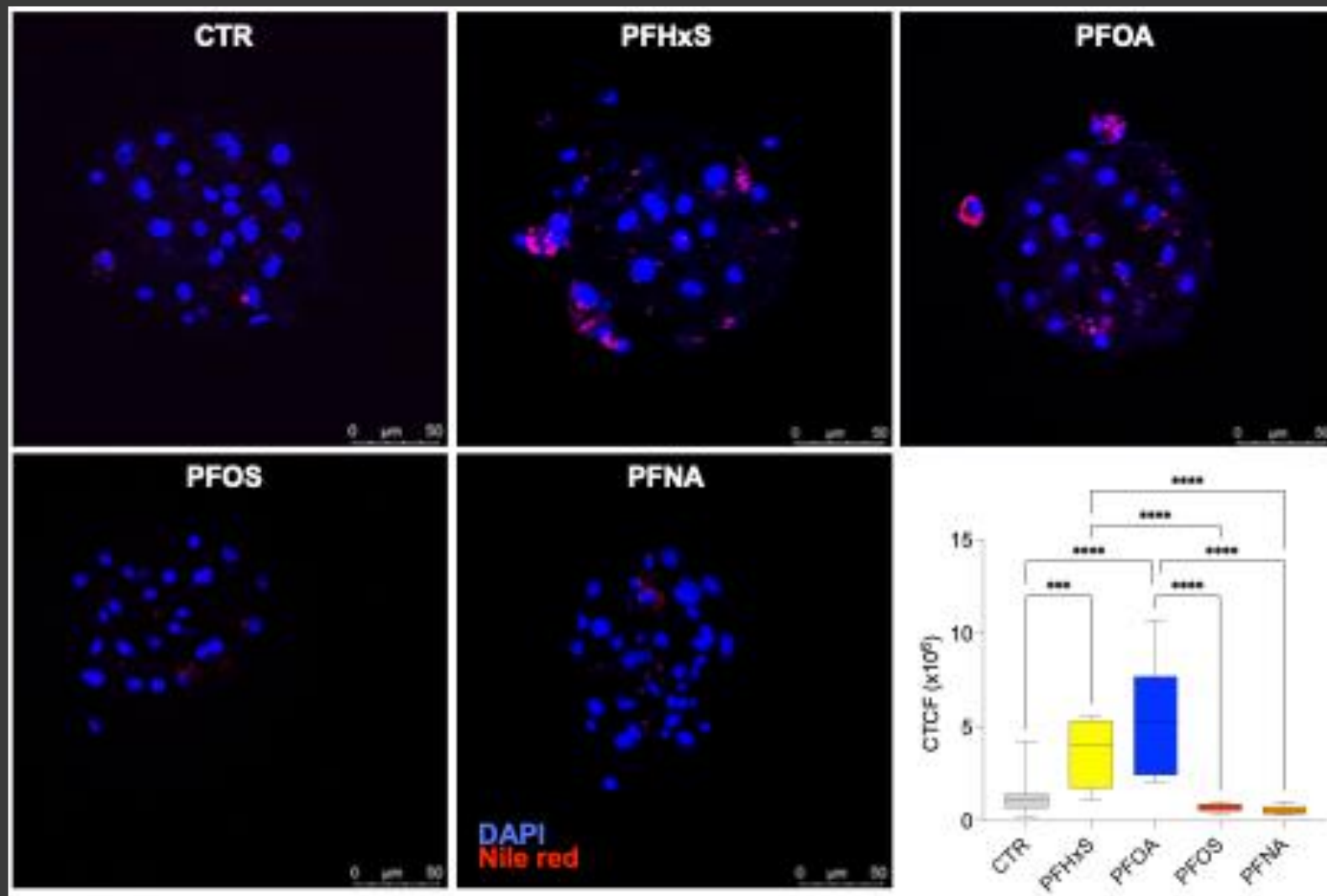
Ana Maretti-Mira, PhD
Co-I – Project 1



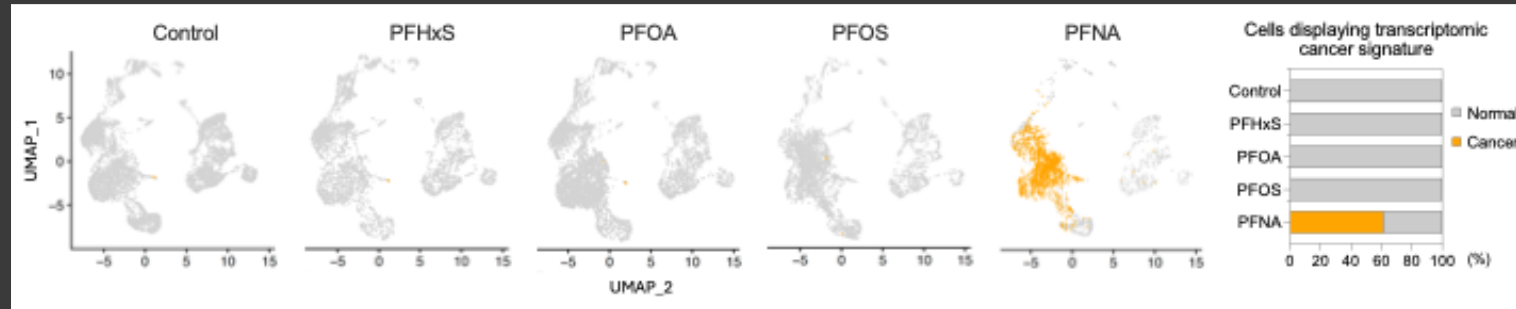
Matthew Salomon, PhD
Co-I – Project 1 and
DMAC



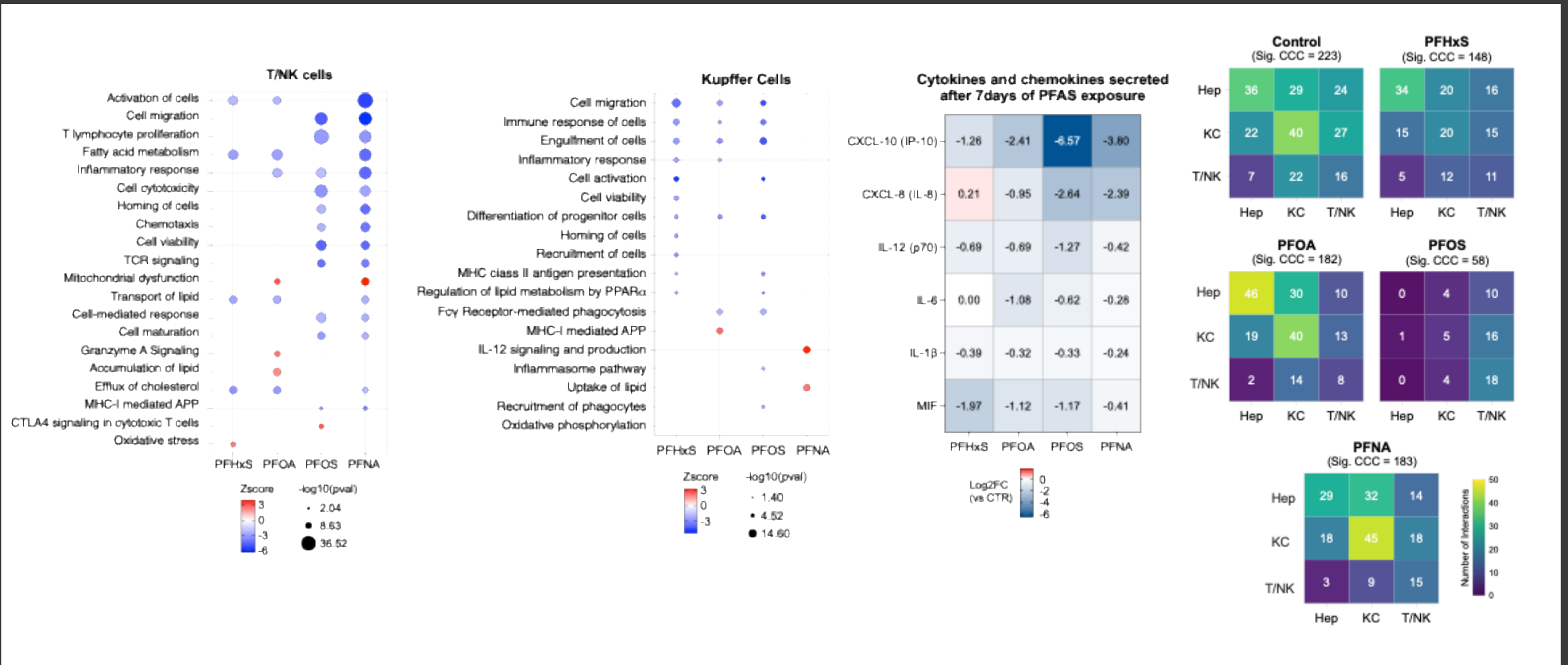
In Vitro Study: Mechanisms of PFAS disruption of lipid metabolism



In Vitro Study: Pro-oncogenic mechanisms of PFAS



In Vitro Study: Immunosuppressive and Cell-Cell Communication Effects



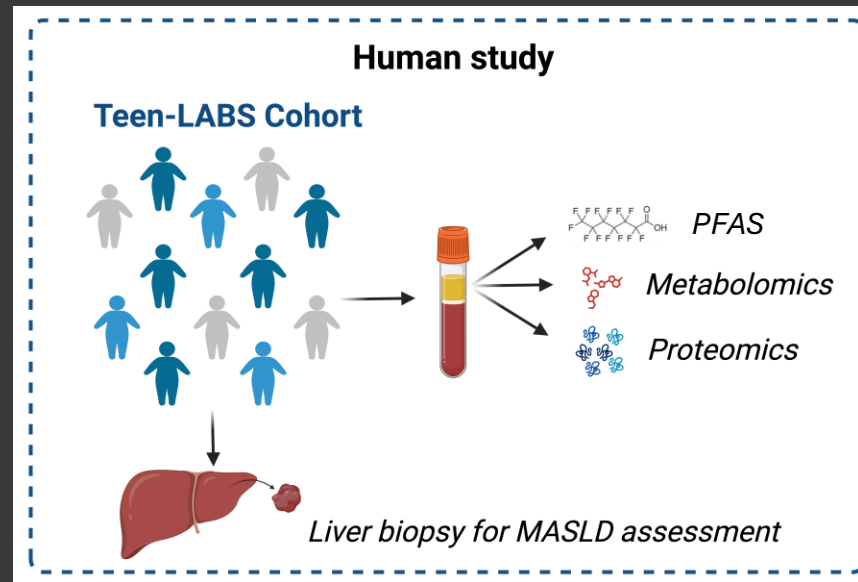
Translational Study: Mechanisms underlying PFHpA-associated MASLD



Lida Chatzi, MD, PhD
ShARP Director
PI – Project 2



Brittney Baumert, PhD
Postdoctoral Scholar
USC



Translational Study: Human Study

Teen-Longitudinal Assessment of Bariatric Surgery (LABS) cohort

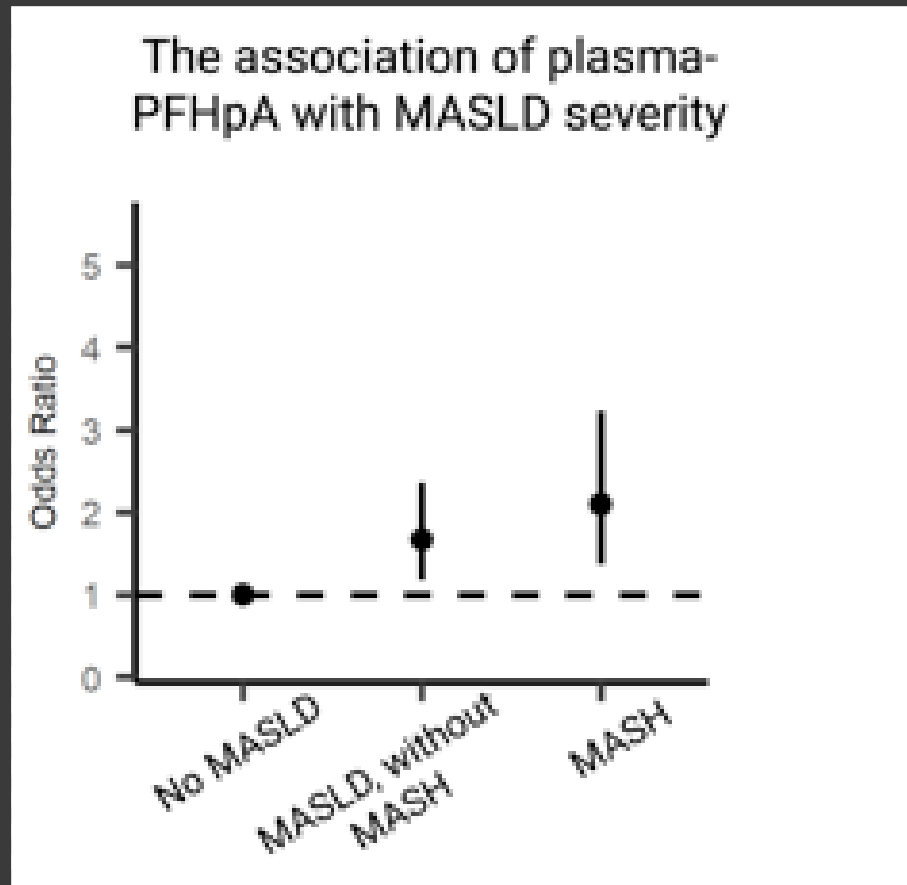
N=186

Age: 17.1 years

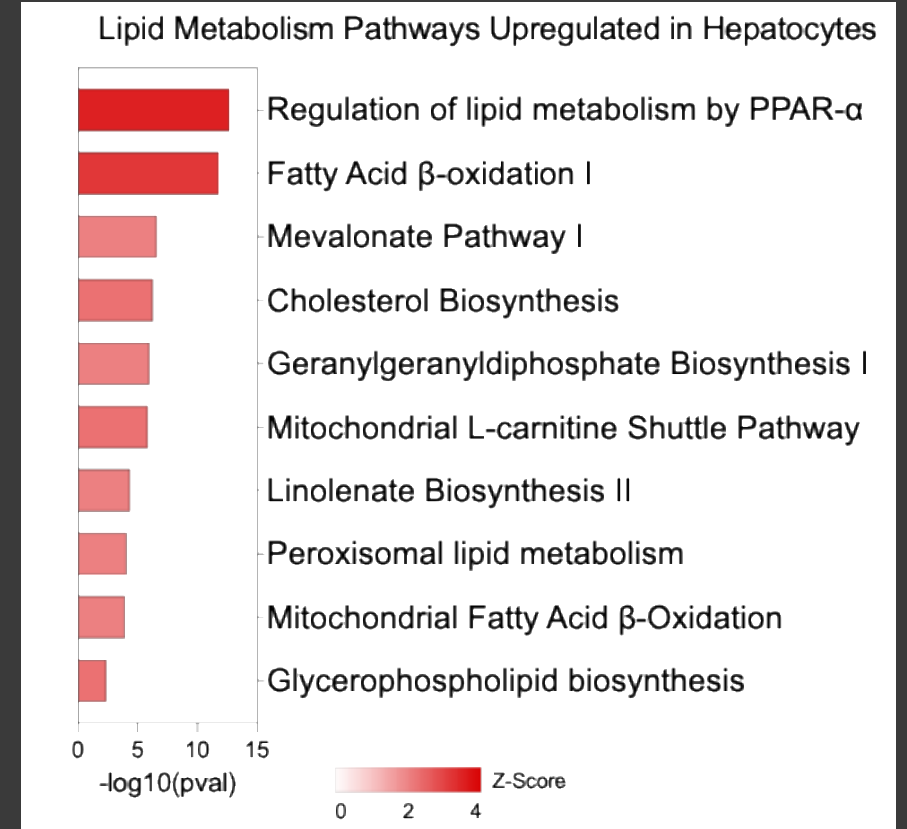
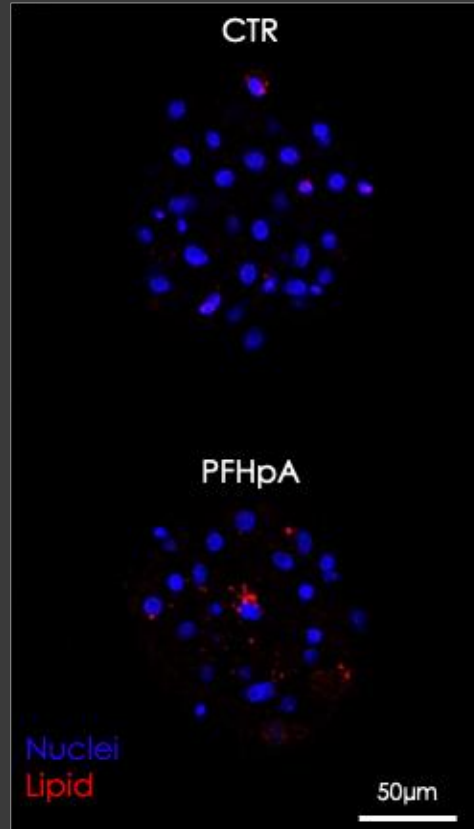
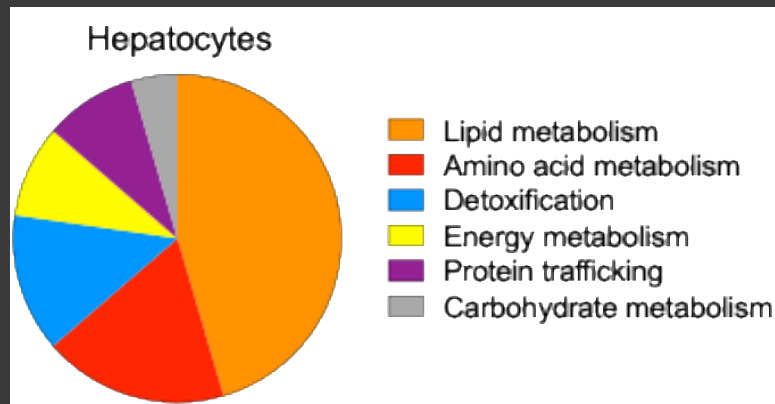
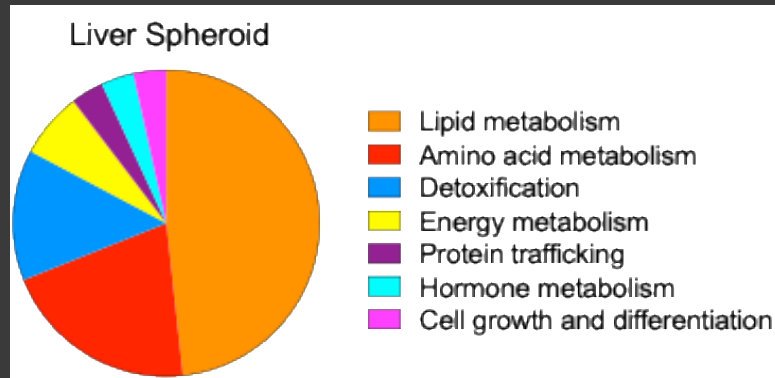
Sex: female (76.3%)

Race: white (72%)

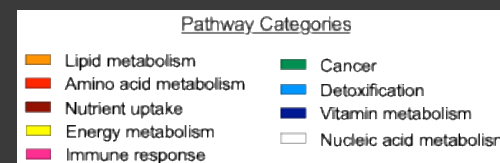
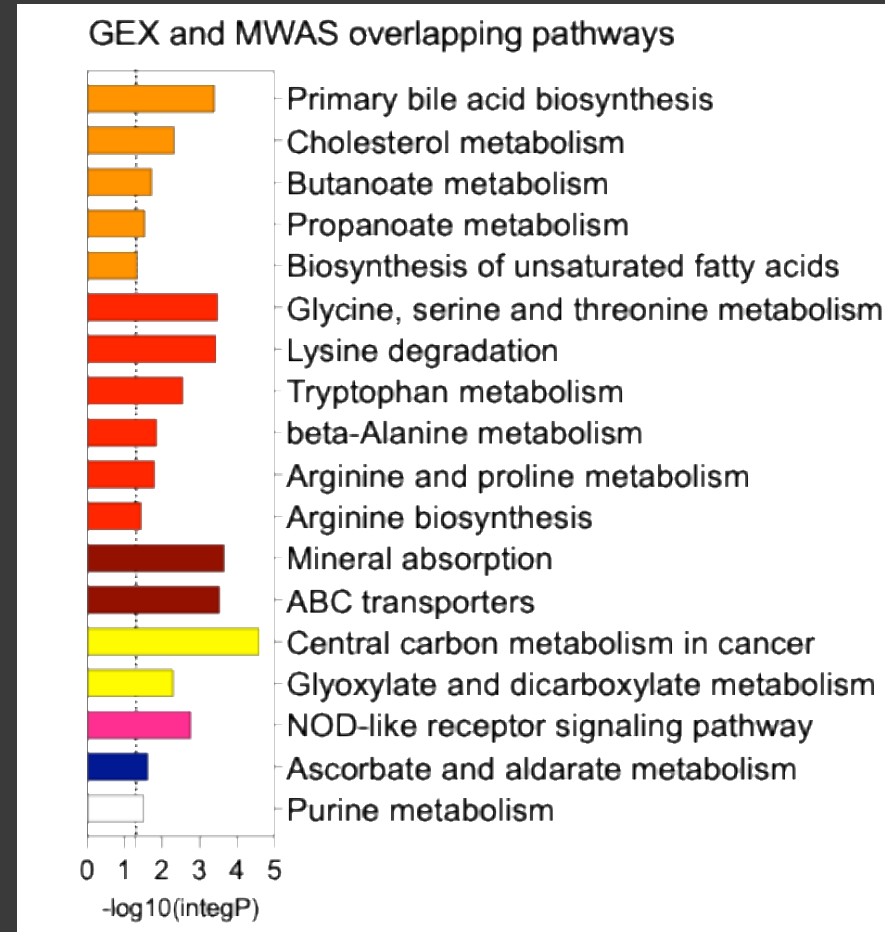
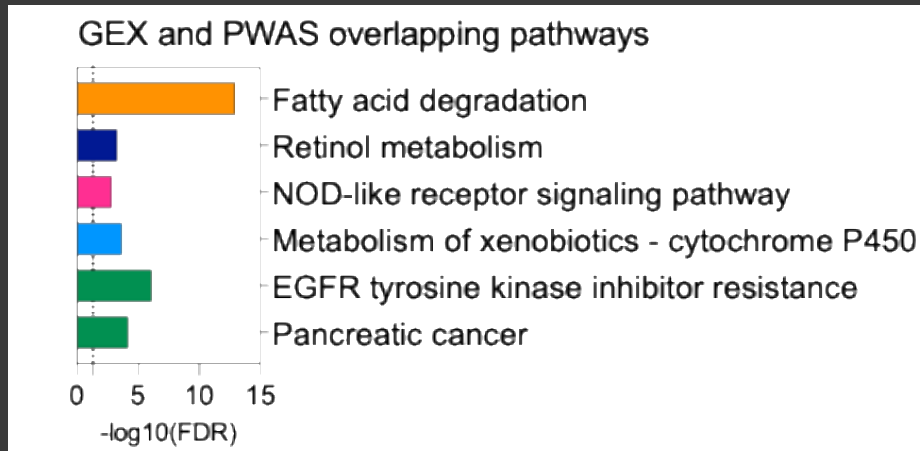
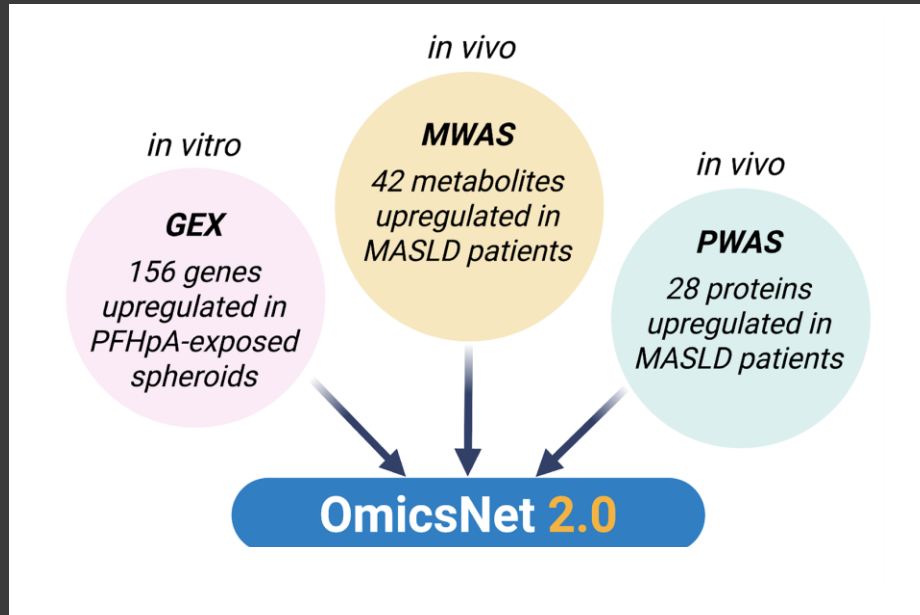
Parental income: \leq \$75k (77.4%)



Translational Study: In Vitro Study



Translational Study: Integration of In Vitro and Human Datasets

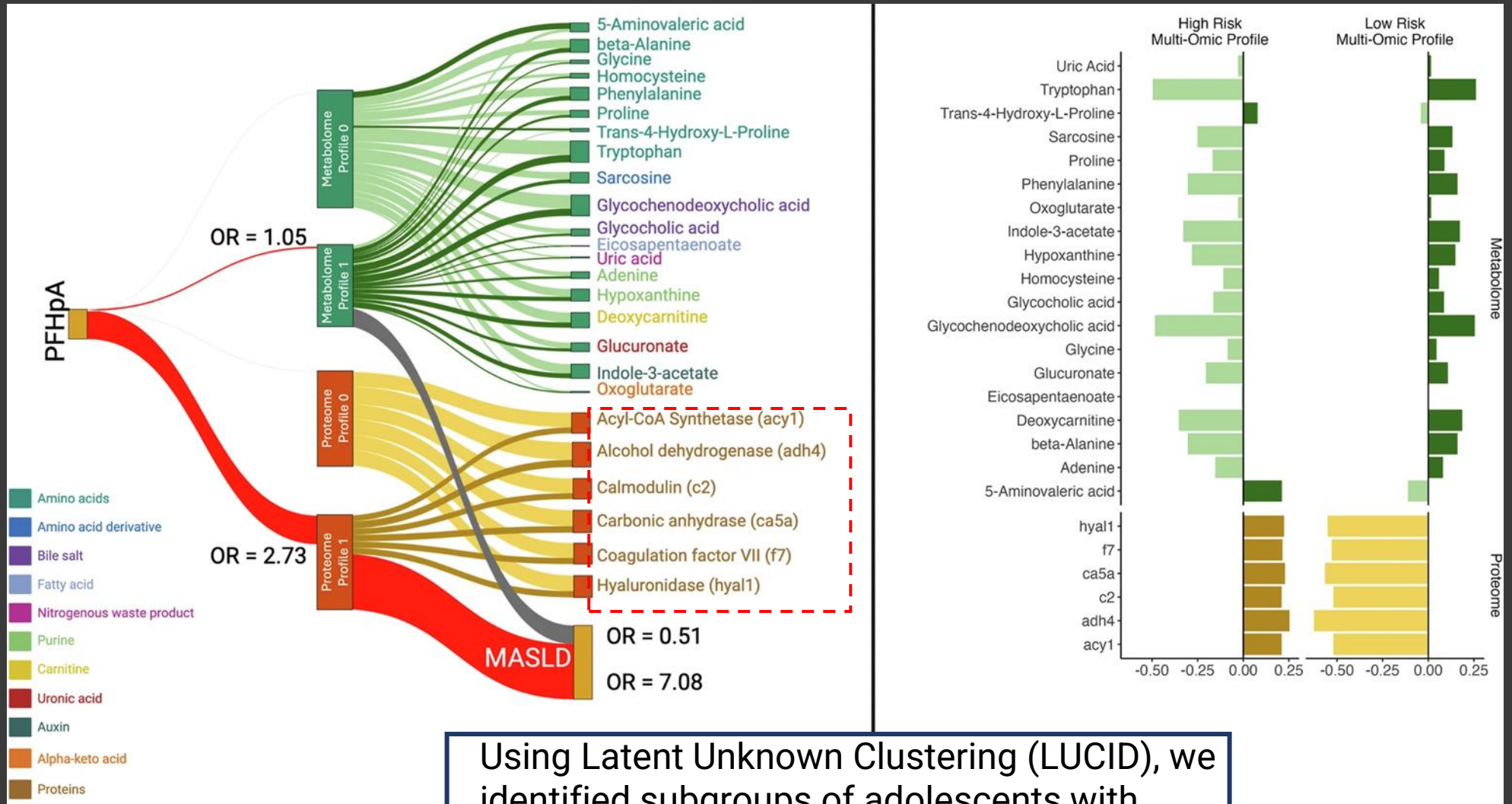


19 metabolites
6 proteins

Translational Study: Extrapolation of In Vitro-Human Data Integration



David Conti, PhD
DMAC Lead



Using Latent Unknown Clustering (LUCID), we identified subgroups of adolescents with increased risk of MASLD

In Progress & Next Steps:

PFAS Mixture
equal amounts of
PFOA, PFOS, PFNA
PFHxS, PFHpA

Final
Concentration
20µM

Exposure Time
Short: 1 week
Long: 2 weeks

Removal
PFAS mix: 1 week
Recovery: 1 week

PFAS Mixture
plasma ratios of
PFOS, PFOA, PFNA
PFHxS, PFHpA
(56:18:6:18:1)

Final
Concentrations
20nM and 80nM

Exposure Time
1 week
3 weeks

Single PFAS
PFOS, PFOA, PFNA
PFHxS, PFHpA

Final
Concentrations
20nM and 80nM

Exposure Time
1 week
3 weeks

Emerging PFAS
6:2FTSA, PFHpS,
TFA

Dose-response
Concentrations
0.02-20µM

Exposure Time
1 week

Acknowledgements

University of Southern California

- Lucy Golden, PhD
- Matt Salomon, PhD
- Lida Chatzi, MD, PhD
- Brittney Baumert, PhD
- David Conti, PhD
- Rob McConnell, MD

Emory University

- Douglas Walker, PhD

UCI

- Veronica Vierra, PhD
- Scott Bartell, PhD

InSphero Inc

- Sue Grepper
- Radina Kostadinova

Funding Sources

P42ES036506;

5P30ES007048

USC Center for Translational Exposomics Research





Website



sharpcenter.usc.edu

Instagram



[@uscsharp](https://www.instagram.com/uscsharp)

LinkedIn



[linkedin.com/company/sharpcenter](https://www.linkedin.com/company/sharpcenter)

Questions?

Ana Maretti-Garcia, MS, PhD

marettim@usc.edu