



## **Welcome to the CLU-IN Internet Seminar**

### **Contaminated Sediments: New Tools and Approaches for in-situ Remediation - Session IV**

Sponsored by: National Institute of Environmental Health Sciences, Superfund Research Program

February 14, 2011, 2:00 PM - 4:00 PM, EST (19:00-21:00 GMT)

*Instructors:*

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*Dr. Harold D. May, Professor, Microbiology and Immunology, Medical University of South Carolina  
(mayh@musc.edu)*

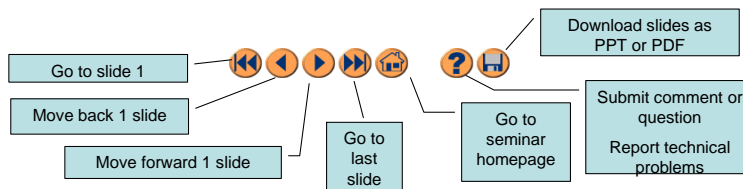
*Moderator:*

*Steve Mangion, U.S. EPA Region 1 (mangion.steve@epa.gov)*

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# Housekeeping

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- Q&A
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- This event is being recorded
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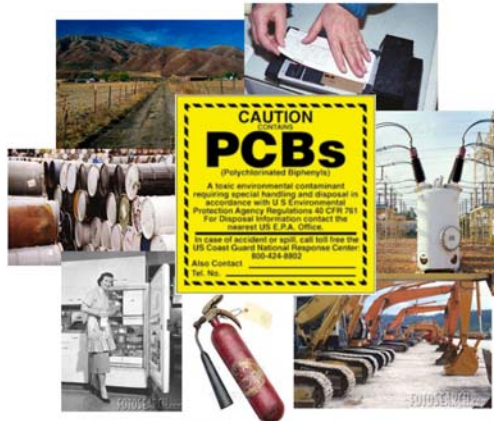
***Integrating Microbial Biostimulation  
and Electrolytic Aeration to Degrade POPs***

**Harold D. May, Kevin R. Sowers  
Chanlan Chun and Ray Payne**

**Medical University of South Carolina  
And  
University of Maryland**

**February 14, 2011**

## ***Persistent Organic Pollutant: Polychlorinated Biphenyls***



Environmental Legacy: Estimated 0.6-1.2 billion kg worldwide

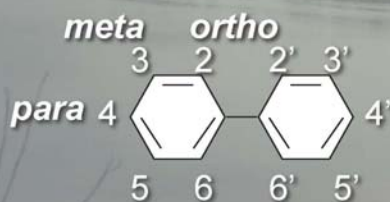
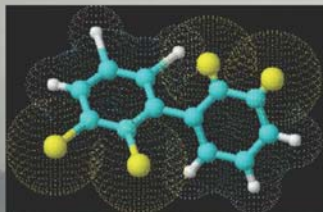
**Goal:**

***Microbial Degradation of PCBs  
in Aquatic Sediment***

**Approach:**

- 1. Identify PCB Dechlorinating Bacteria***
- 2. Develop in situ Monitoring Tools***
- 3. Develop Bioaugmentation***
- 4. Test Bioelectrochemical Stimulation***

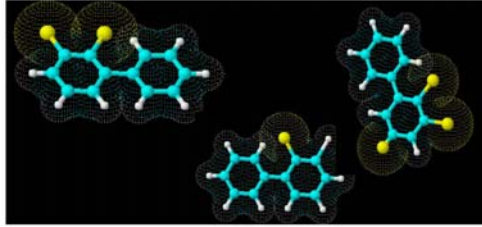
## Polychlorinated Biphenyls (PCBs)



- 209 congeners
- Very stable
- Bioaccumulate
- Toxicity concern
- Anoxic sediments are sinks

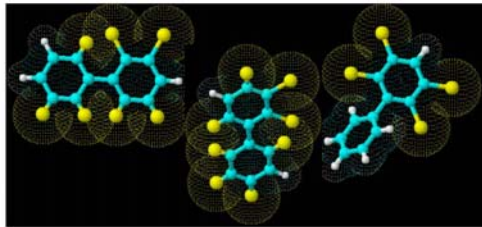


## ***Microbial PCB Degradation***



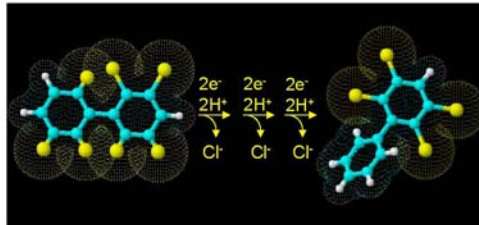
Complementary processes

✓ Aerobic biodegradation  
Generally with < 4-6 Cl



✓ Anaerobic dechlorination  
generally with  $\geq 3$  Cl

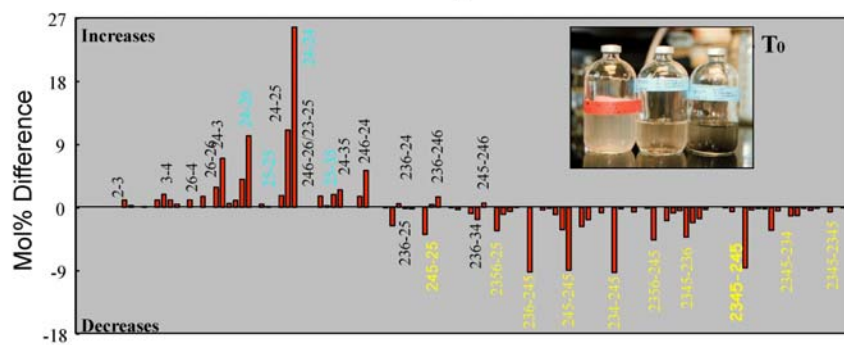
## ***Reductive Dechlorination of PCBs***



- ✓ Performed by anaerobes, e.g. members of the Chloroflexi such as strains DF1, SF1 and *Dehalococcoides* strain CBDB1
- ✓ Theoretically can produce biphenyl but rarely do. Ordinarily leave mono, di and trichlorobiphenyls behind



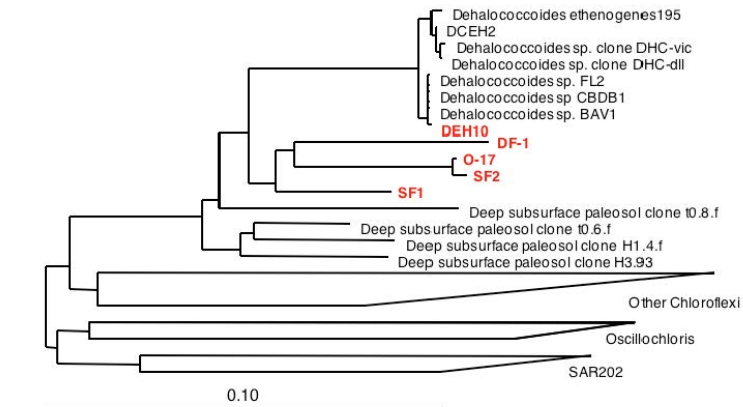
## Dechlorination of Aroclor 1260



Wu, et al. *AEM* 1998



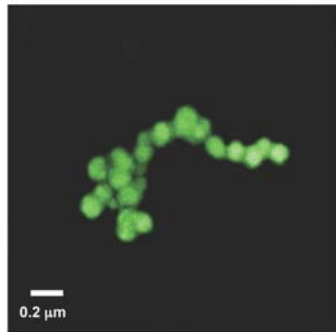
## PCB Dechlorinating Bacteria



Based on 16S rRNA gene sequences (1500+ bp)

Cutter et al. *Environ. Microbiol.* 2001, Wu et al.,  
*AEM* 2002, Fagervold et al. *AEM* 2006 & 2007,  
 Kjellerup et al. *AEM* 2008

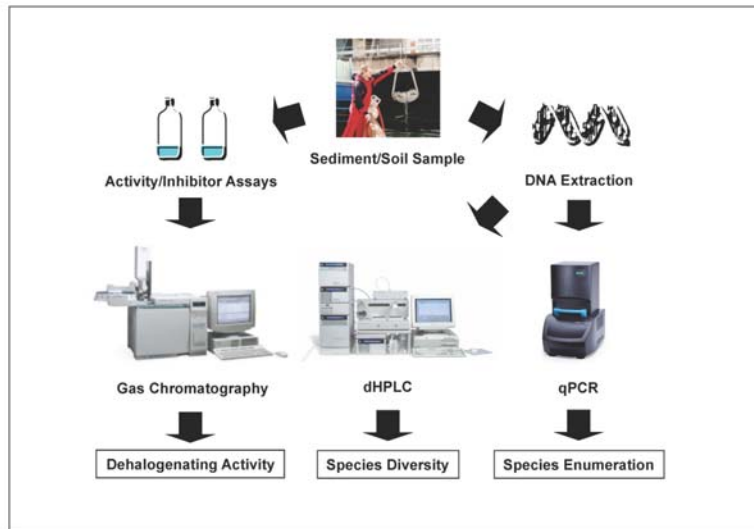
## *Detecting PCB Dechlorinating Bacteria*



- Optical density
- Plating
- Direct cell counts
- FISH

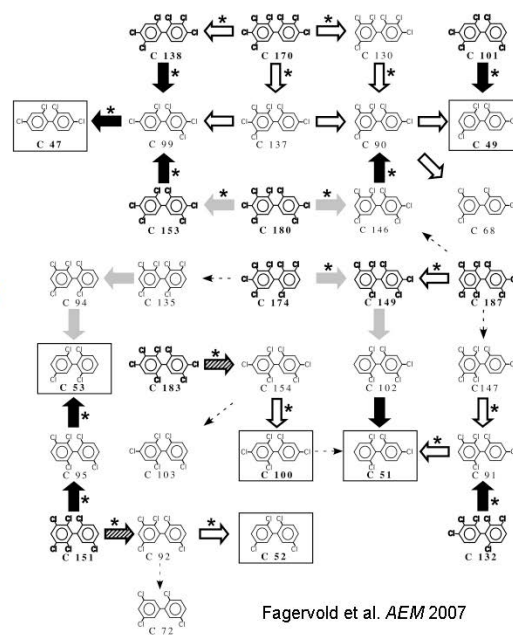
May et al, 2008

## *High Throughput Microbial Analysis/Monitoring*



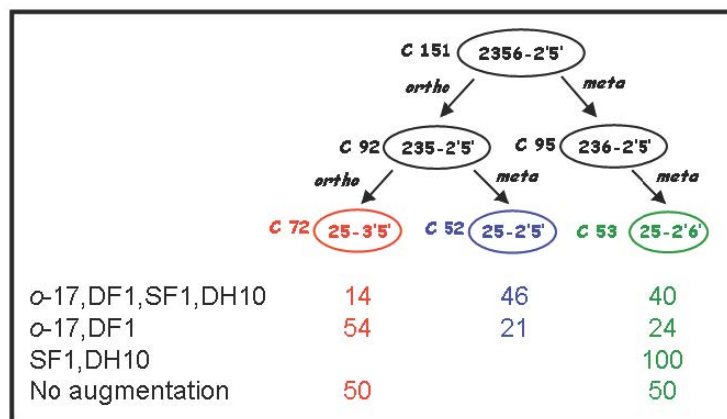
## Dechlorination of Congeners Predominant in Aroclor 1260

Only 3 Phylotypes Required



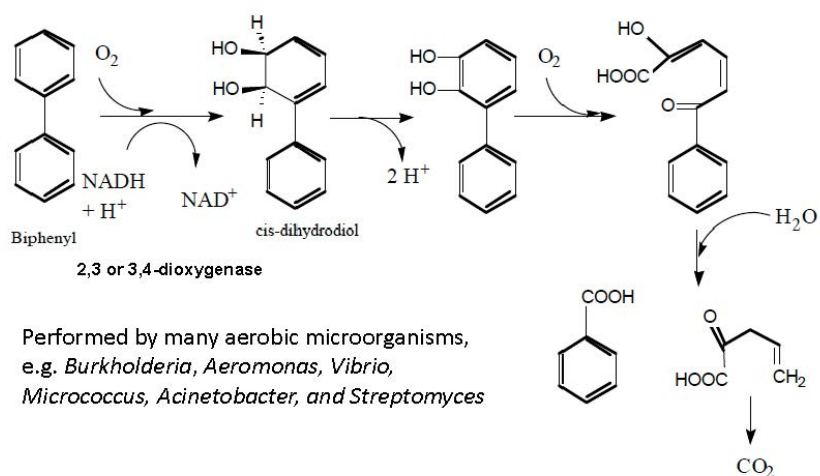
Fagervold et al. AEM 2007

## Bioaugmentation with PCB Dechlorinators

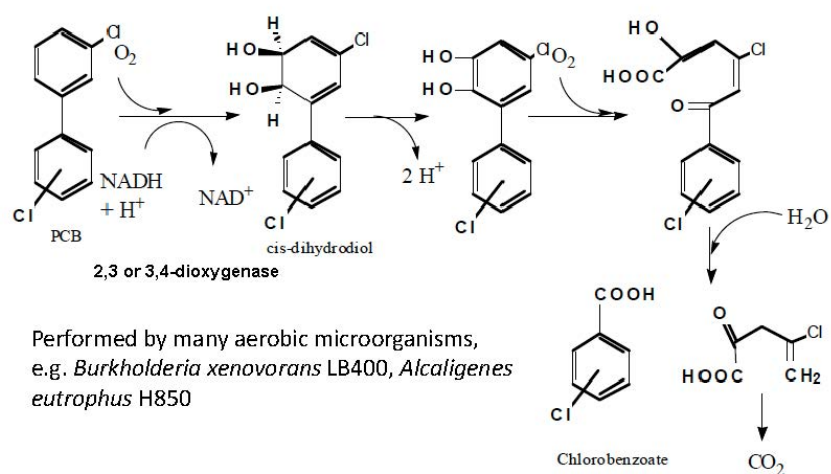


Fagervold, submitted

## Aerobic Biodegradation of Biphenyl



## Aerobic Biodegradation of PCBs





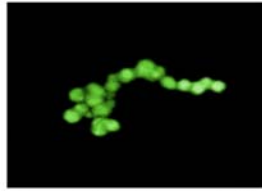
## ***Bioaugmentation of weathered Aroclors***

Baltimore Harbor Sediment: 5-10 ppm weathered PCBs

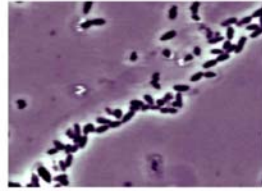


- 1 liter sediment incubated at 20°C in dark
- indigenous water with no additional nutrients
- PCB dechlorinating anaerobe DF1  
and/or PCB degrading aerobe LB400

## ***Bioaugmentation of weathered Aroclors***



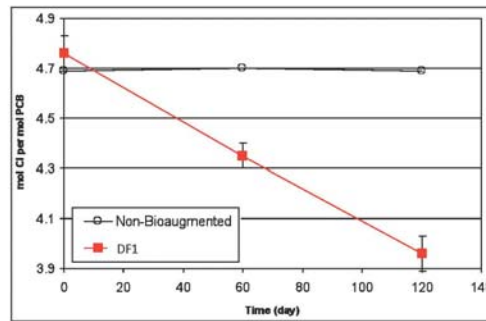
*Dehalohalobium chlorocoercia* DF-1



*Burkholderia xenovorans* LB400

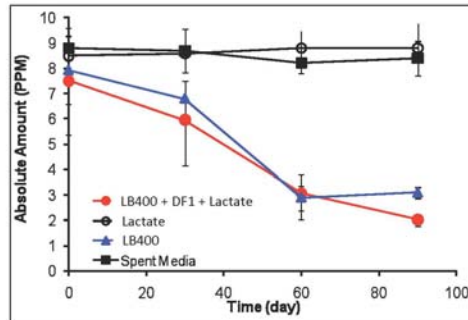
- each mesocosm was inoculated with  $5 \times 10^5$  cells
- inoculation by direct injection or on solid substrate
- lactate added as carbon source when indicated

## ***Bioaugmentation of Baltimore Harbor Sediment with DF1***



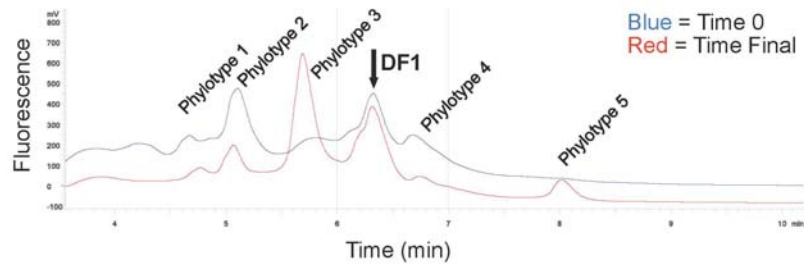
DF1 removed about 0.9 chlorines per mol PCB in 120 days.

## ***Bioaugmentation with DF-1 + LB400 Results in Degradation of Weathered PCBs***

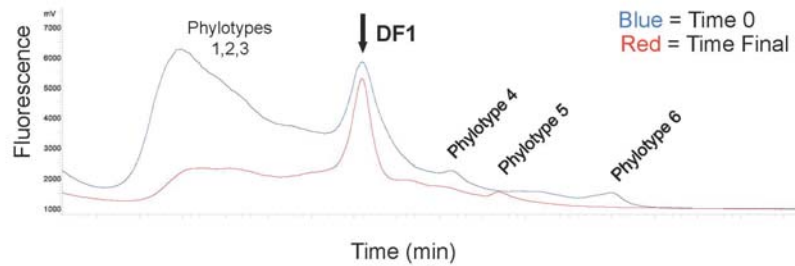


About 75% of total PCBs by mass degraded through oxidative processes and reductive dechlorination in 90 days. Experiment ongoing.

## Dechlorinator phylotypes in mesocosm amended with DF1

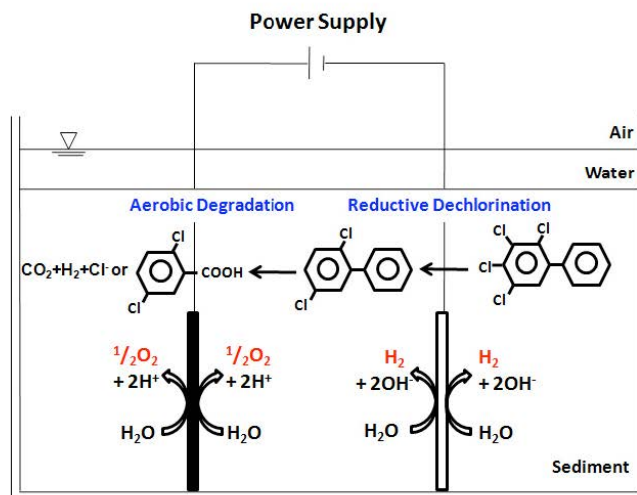


### Dechlorinator phylotypes in mesocosm amended with DF1 & LB400



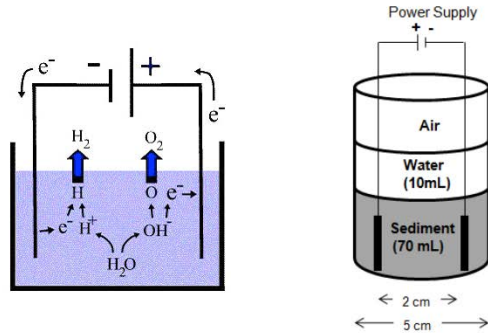
- DF-1 detected by dHPLC at similar levels on days 0 and 90
- Fate of LB400 currently being determined

## Bioelectrochemical Approach: Hypothesis



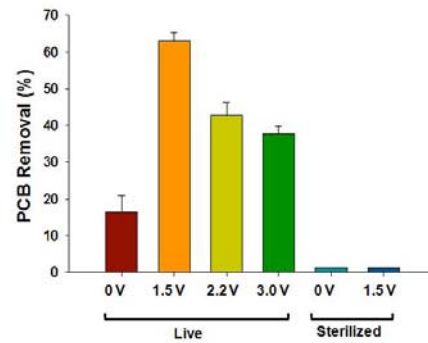
## Sediment Bioelectrochemical Reactors (SBRs)

- Microcosm-SBRs
  - Aroclor 1242 impacted sediment (~20 ppm) from Fox River, WI
  - Electrode: Ti sheet (surface area:  $12.5 \text{ cm}^2$ )
  - Applied voltage: 0, 1.5, 2.2, and 3 V



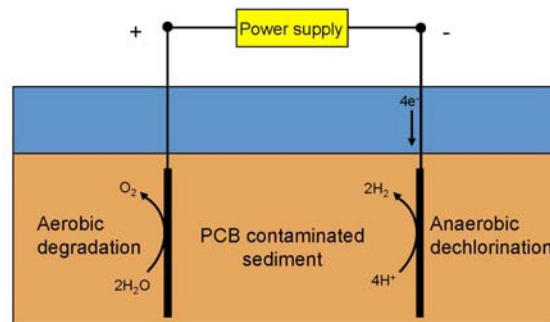


## Sediment Bioelectrochemical Reactors (SBRs)



Up to 65% of total weathered PCBs by mass degraded through oxidative processes in 88 days without bioaugmentation. Increasing chlorobenzoates and benzoate detected with all voltages. Analysis continues.

## *Application of Voltage to Aquatic Sediment*



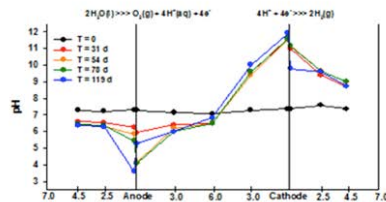
pH problem?

## Problem with pH

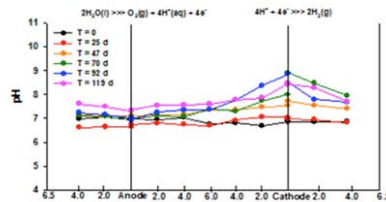
(Redox,  $O_2$  and  $H_2$  gradients follow suit)

High current density : 0.067 mA/cm<sup>2</sup> (3.8-4.4 V, 15.2-17.6 mW)      Low current density : 0.003 mA/cm<sup>2</sup> (2.0-2.5 V, 0.036-0.045 mW)

Electrode potential vs. Ag/AgCl  
Cathode: -1.6 V ( $H_2$  generation)  
Anode : 2.5 V ( $O_2$  generation)



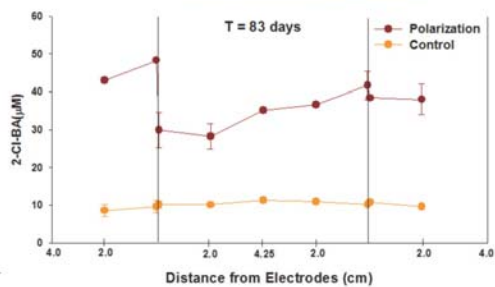
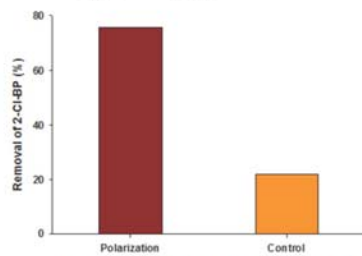
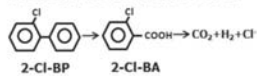
Electrode potential vs. Ag/AgCl  
Cathode: -1.1 V ( $H_2$  generation)  
Anode : 1.4 V (no or slow  $O_2$  generation)



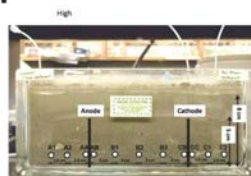
## SBR-Polarization

- Mesocosm-SBR

- PCB-impacted sediment from Fox River, WI
- Electrodes: Ti sheet (surface area: 55cm<sup>2</sup>)
- Constant Current: 0.014 mA/cm<sup>2</sup>, 2.5-3.8 V
- Model PCB added: 2-Cl-BP



Repeated polarization (cycling of the potential) avoids pH shifts and supports PCB degradation throughout the mesocosm. Weathered PCBs to be tested.



### ***Conclusions***

- Bioaugmentation results in 75% PCB degradation in 90 days
- Application of 1.5V results in 65% PCB degradation
- Anaerobic oxidation of PCBs is hypothesized
- Repeated polarization is successful and avoids pH problems

### ***Future Goals***

- Determine full extent of effect of bioaugmentation and electrochemical stimulation
- Determine extent of repeated polarization
- Combine bioaugmentation and electrochemical stimulation
- Test *in situ*



Danny Reible

Environmental and Water Resources  
University of Texas

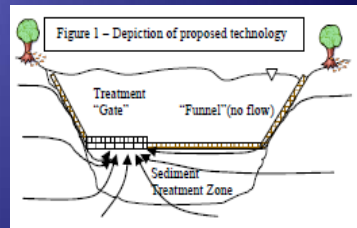
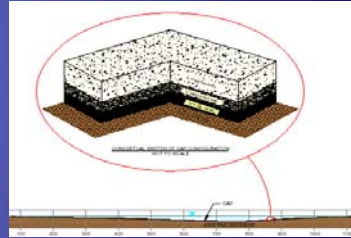
## FUNNEL AND GATE APPROACH FOR ACTIVE SEDIMENT CAPS

CO-INVESTIGATORS:

G. LOWRY, K. GREGORY, CARNEGIE-MELLON  
J. HUGHES, GATECH

# Sediment In-situ Capping/Treatment

- ◆ Reduce risk by:
  - Stabilizing sediments
  - Physically isolating sediment contaminants
  - Reducing contaminant flux to benthos and water column
- ◆ Sand effective for strongly solid associated contaminants
- ◆ “Active caps” for other situations (w/amendments)
- ◆ Funnel and Gate approach to maximize ability to place/replace amendments

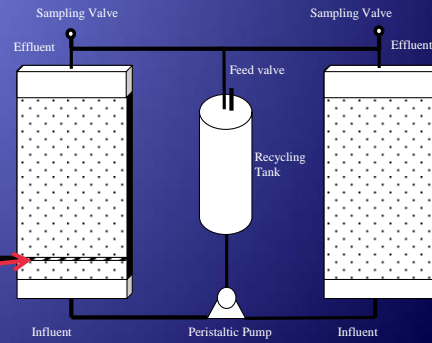
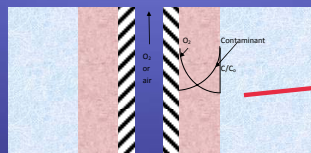
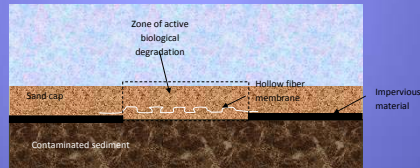


## Project Objectives and Scope

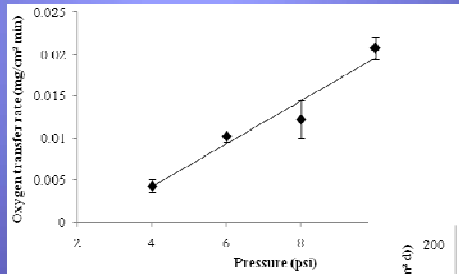
- ♦ Can a funneling cap effectively contain contaminants and effectively channel interstitial fluids (water or NAPL) to a collection or treatment gate
- ♦ Can treatment gates composed of reactive materials effectively manage residual contamination from the in-situ treatment zone
- ♦ Scope
  - Preliminary studies with sorbents (e.g. organoclays and activated carbon)
  - Hollow fiber membrane to introduce oxygen in reduced environment
  - Low voltage electrodes to encourage development of appropriate redox conditions



# Hollow fiber membrane to introduce oxygen & encourage aerobic degradation



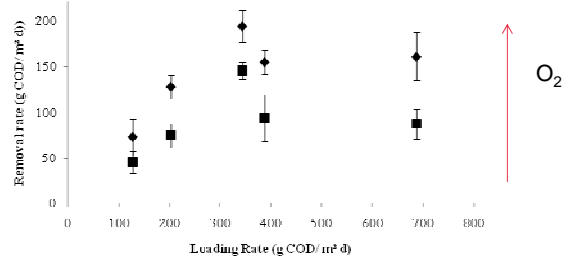
# Hollow Fiber Membrane Induced Oxidation



Initially reduced conditions without oxygen introduction via pressurized air

Minimal membrane fouling

Acetate oxidation



## The idea of electro-reactive capping

- ◆ Functions:

- Provide redox control
- Direct reduction and oxidation on electrode surface
- Deliver electron donor and acceptor for further degradation

- ◆ Advantages:

- Real time and site-specific control
- complete mineralization of contaminants through sequential reduction and oxidation

## The idea of electro-reactive capping

- ◆ Functions:

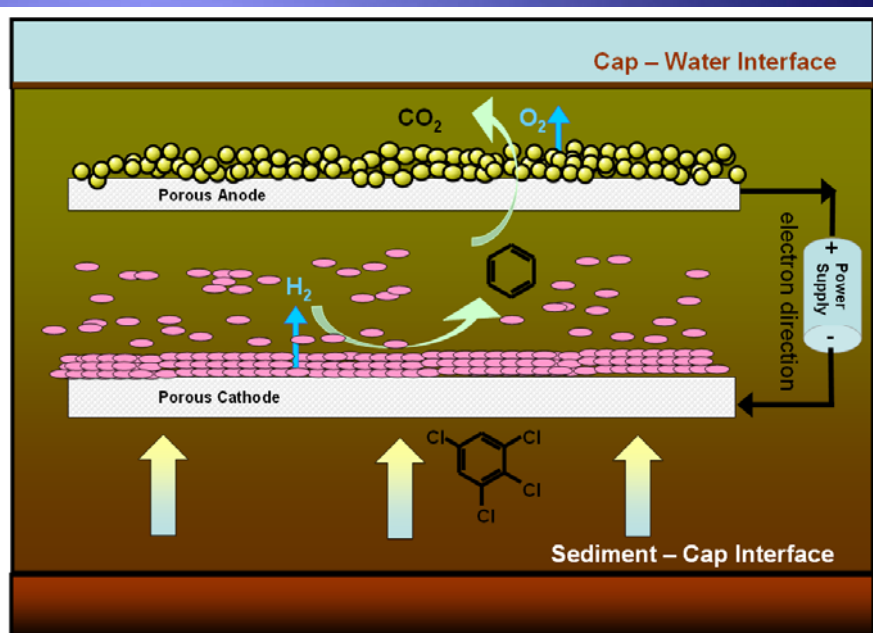
- Provide redox control
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- ◆ Advantages:

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- complete mineralization of contaminants through sequential reduction and oxidation



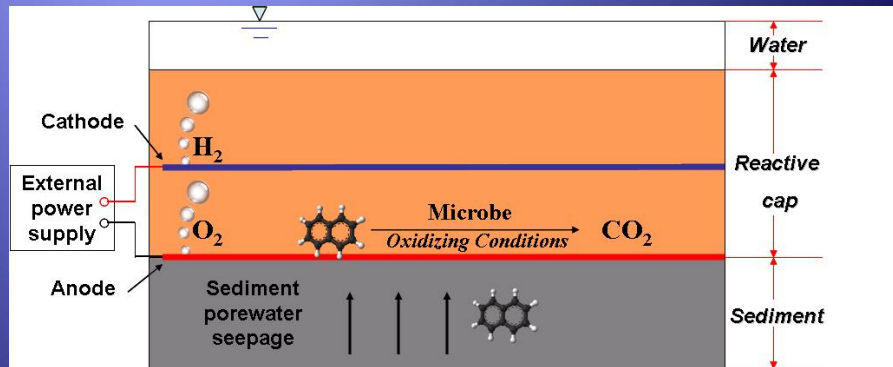
## Conceptual model- Electrode cap



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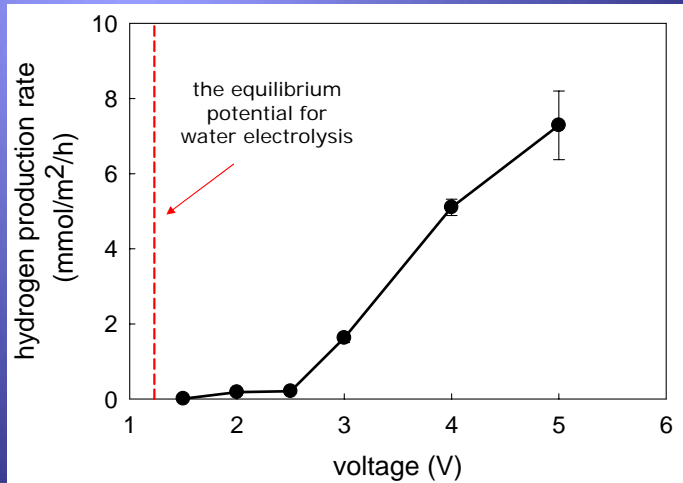
# Configuration for PAH oxidation

- ◆ Anode at depth to encourage oxidizing conditions where normally strongly reducing conditions exist



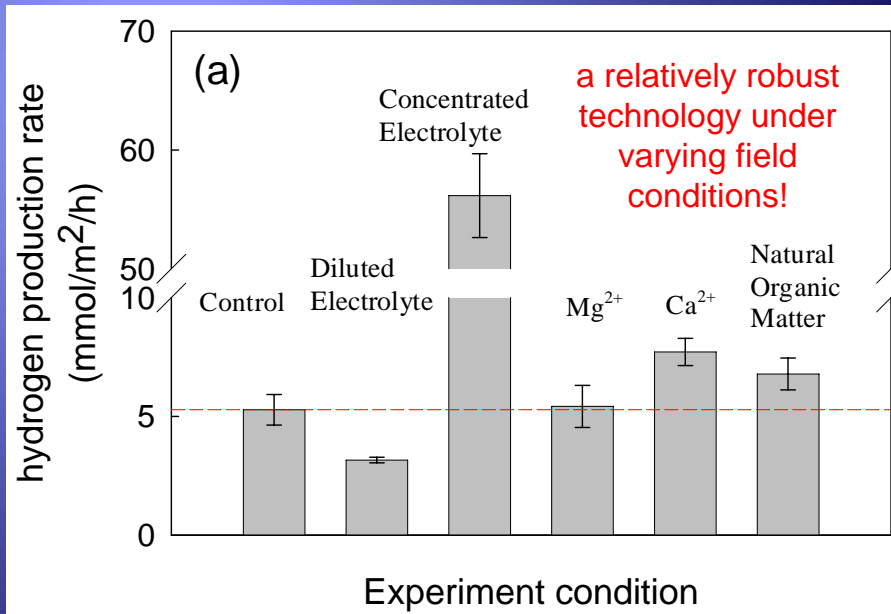
## Influence of voltage

- ◆ Real-time control of electron donor production in the cap by adjusting voltage



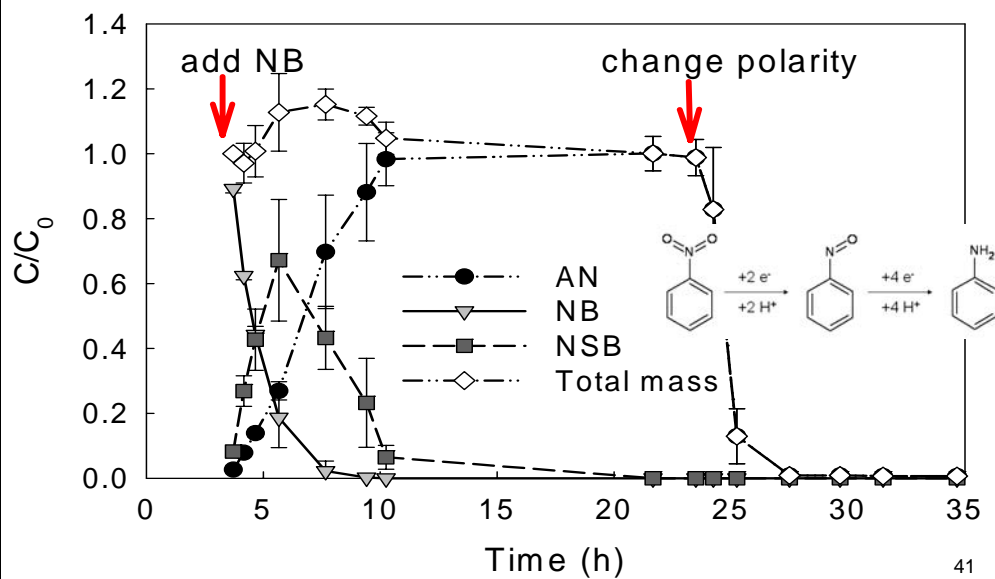
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## Influence of aqueous chemical species

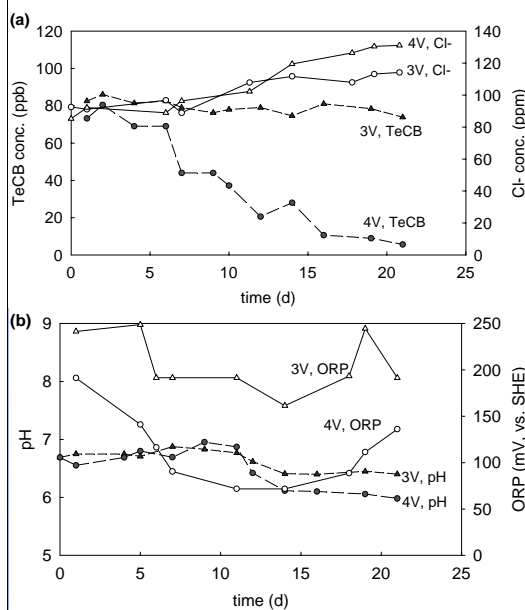




## NB sequential reduction-oxidation



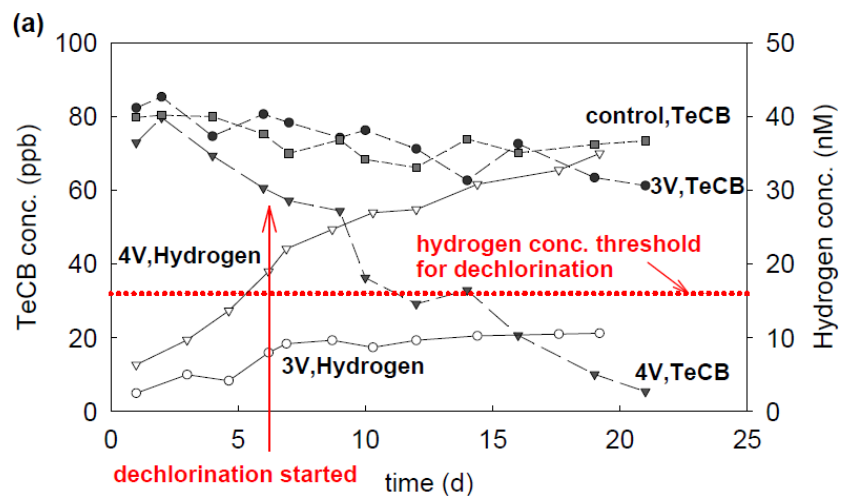
# Electrode stimulated degradation of 1,2,3,5-Tetrachlorobenzene



3v insufficient  
4v removed ~90% TeCB

ORP: from 200mv to 70mv  
pH changes: less 1 unit

# Electrode induced dechlorination

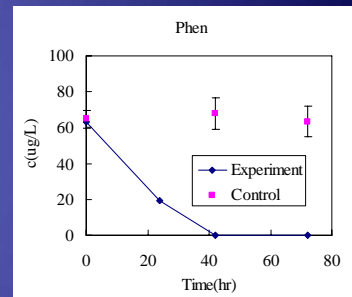
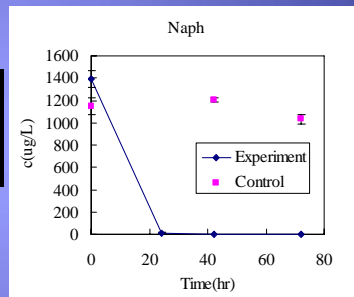


# PAH biodegradation in slurry

- ♦ PAH degradation under aerobic and nitrate reducing condition
  - To verify PAH biodegradation potential by indigenous microbe in sediment
  - To prove that biodegradation rate is faster in more oxidizing condition (aerobic condition)
- ♦ Electrode enhanced degradation of PAH in slurry
  - To examine the feasibility of electrode enhanced biodegradation of PAH in slurry phase

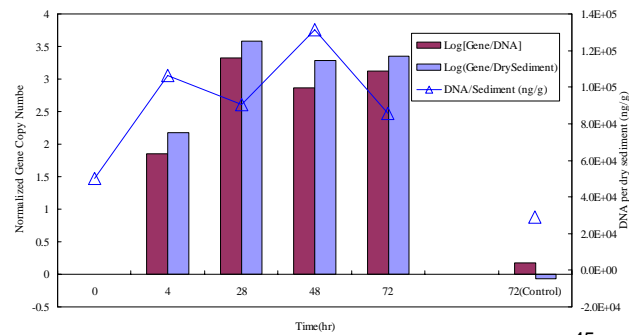
## PAH biodegradation under aerobic conditions

**Naphthalene/  
Phenanthrene  
Concentration**



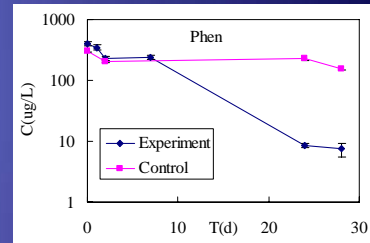
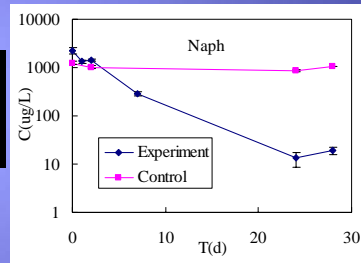
### PAH degrading genes

Quantitative polymerase chain reaction (qPCR) using primers that target PAH ring-hydroxylating dioxygenase (PAH-RHD) genes

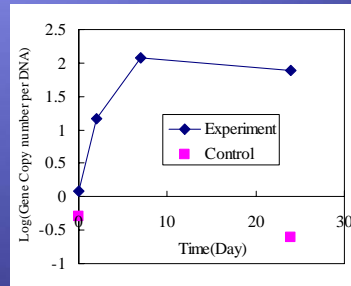


## PAH biodegradation under nitrate reducing condition

**Naphthalene/  
Phenanthrene  
Concentration**



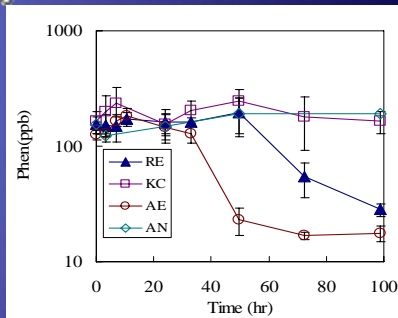
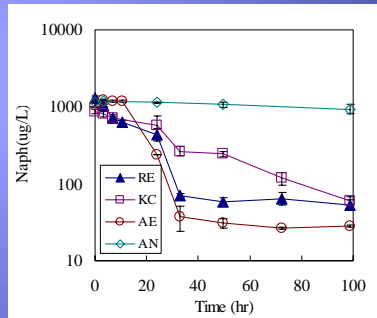
**PAH  
degrading  
genes**



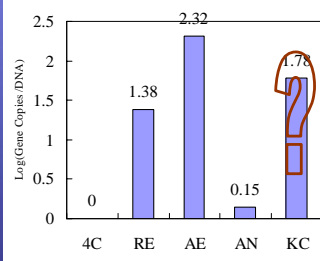
# Electrode enhanced biodegradation of PAH in sediment slurry

Initially anaerobic slurries

Except fully aerobic slurry



PAH degrading genes



Constant condition slurries

AE: Aerobic condition

AN: Anaerobic condition

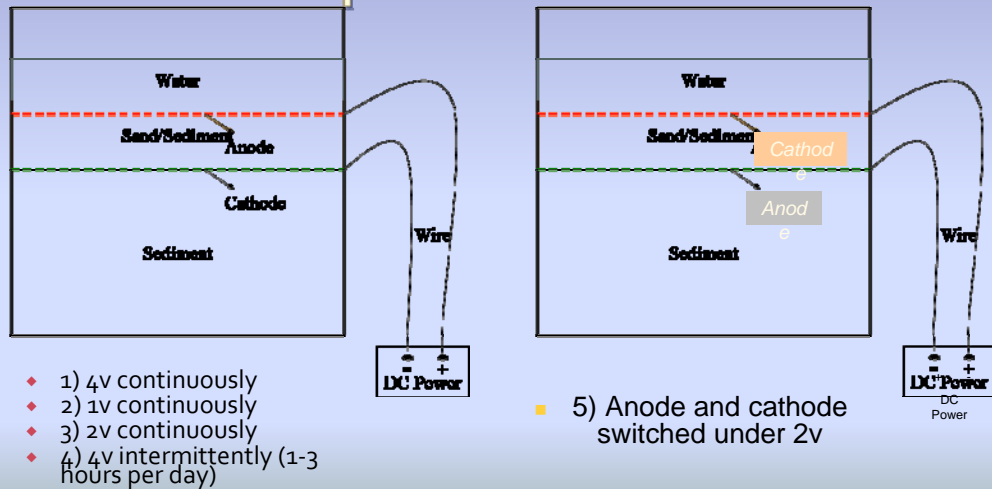
Electrode induced aerobic conditions

RE: Electrode biodegradation

KC: Killed control

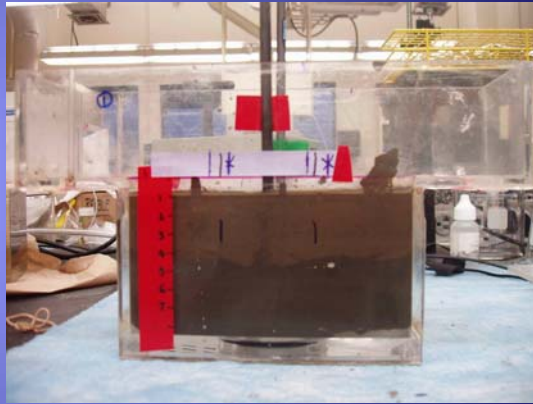


## Redox control and pH changes in sediment caps with electrodes





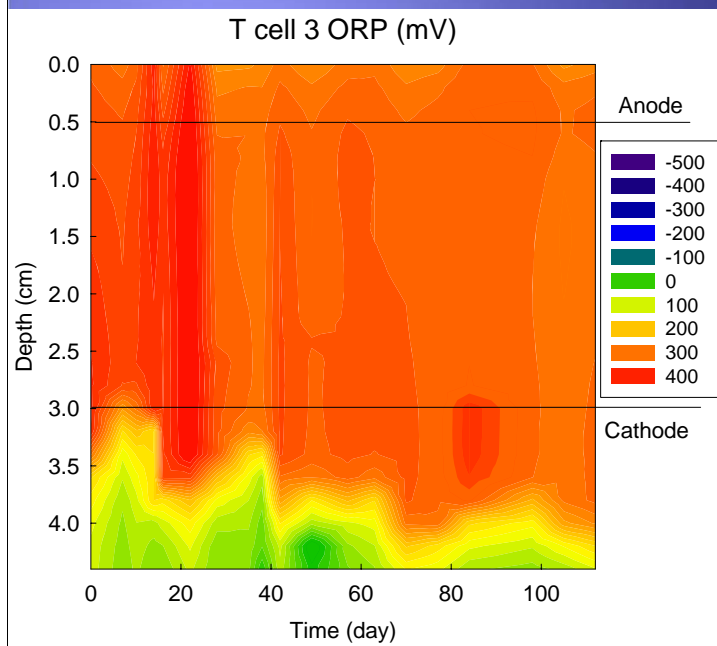
- Side View



- Top View



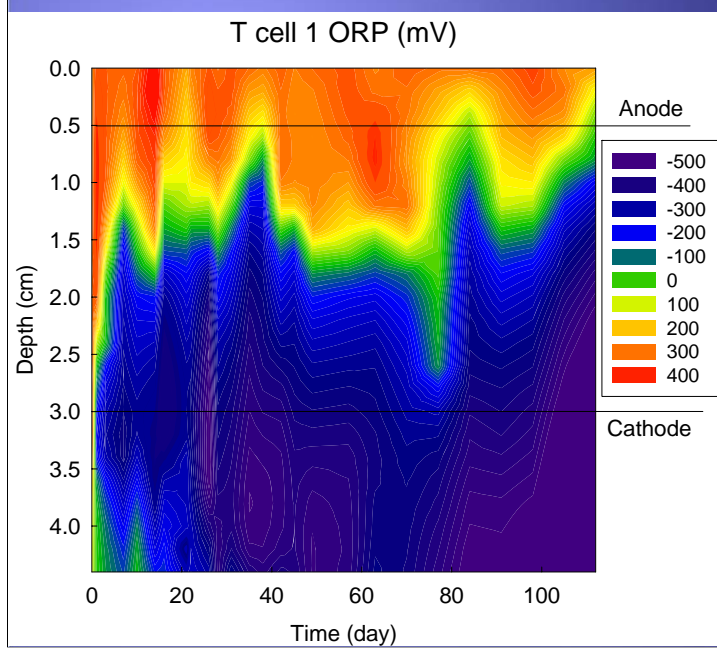
# Redox control



Unpowered  
control

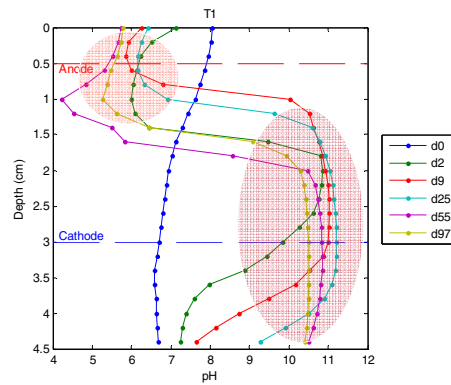
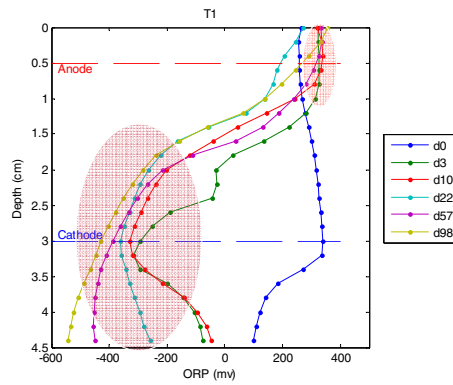
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# Redox control



Continuously  
connected  
to 4V DC  
power supply  
for ~100 days

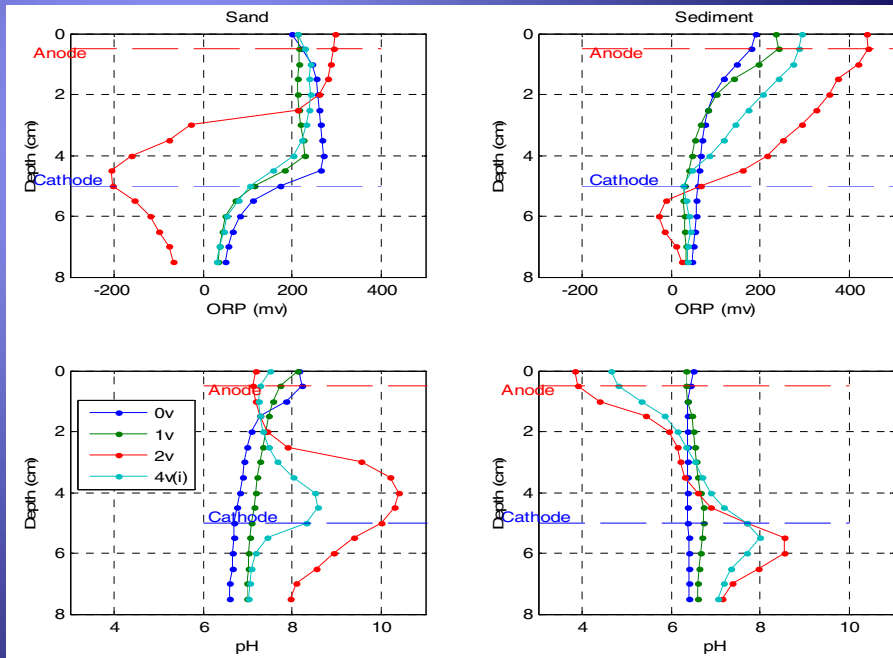
# Redox and pH in 4v experiment



Oxidized zone <-> acidic zone

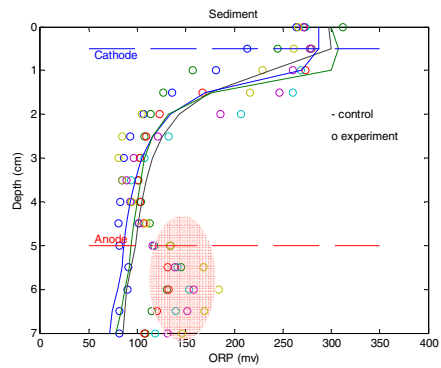
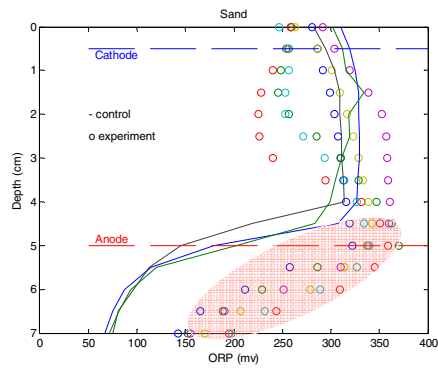
Reduced zone <-> basic zone

## Redox and pH in 1v,2v,4v(intermittently) experiment



# Redox in *deep oxidation mode*

- ◆ Anode at depth (sediment-cap interface)
- ◆ PAH degradation mode



# Redox control by electrodes

- ◆ Proof of concept was successful
- ◆ Electrodes could establish oxidizing conditions at the anode
  - Currently studying degradation and increases in PAH degrading genes as per slurry experiments
- ◆ pH changes were always associated with redox changes
  - Currently studying use of siderite (iron carbonate) as buffer material to control pH

# Intermediate Scale Demonstration

- ◆ Large flume demonstrations in cooperation with Bayani Cardenas (UT Geosciences)
- ◆ 2 dimensional flow (upwelling, river flow, hyporheic exchange)





## Conclusions

- ♦ Funnel and gate approach allows significantly more complex and sophisticated capping
  - Regular replacement of finite capacity sorbents
  - Sustainable biodegradation
- ♦ Hollow fiber membranes to introduce air/oxygen and encourage aerobic degradation
  - Enhanced degradation shown
- ♦ Low power electrodes for encouragement of reduction/oxidation at different layers in sediments
  - Enhanced degradation shown in idealized systems
  - In-sediment studies ongoing
  - Intermediate scale demonstrations planned

# Resources & Feedback

- To view a complete list of resources for this seminar, please visit the [Additional Resources](#)
- Please complete the [Feedback Form](#) to help ensure events like this are offered in the future

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